



Evaluation of antifungal activity of ferrous oxide (Fe_2O_3) and ferric oxide (Fe_3O_4) nanoparticles on *Aspergillus flavus*

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ABSTRACT

Nanoparticle is a new innovation of using particles in a new size which ranged from 1- 100 nanometers that equal 1×10^{-6} meter. In this study we evaluated the antifungal effect of Fe_2O_3 and Fe_3O_4 nanoparticles on *Aspergillus flavus* (KP137700) selected for the study isolated from broiler feed by Mycology Department of the Animal Health Research Institute, Giza, Egypt. Fe_2O_3 and Fe_3O_4 nanoparticles synthesized by co-precipitate method. The size of synthesized nanoparticles was 45 nanometer for Fe_2O_3 and 9 nanometer for Fe_3O_4 . The antifungal effect Fe_2O_3 and Fe_3O_4 nanoparticles against *Aspergillus flavus* measured by MIC method and evaluated by scanning electron microscope. The results revealed that Fe_2O_3 nanoparticles have antifungal effect more than Fe_3O_4 nanoparticles.

Keywords: *Nanoparticles, Fe_2O_3 , Fe_3O_4 , Antifungal, *Aspergillus flavus*.*

(<http://www.bvmj.bu.edu.eg>)

(BVMJ-34(2): 77-83, 2018)

1. INTRODUCTION

Nanotechnology has offered great possibilities in various fields of science and technology (Adibkia *et al.*, 2007). Pharmaceutical nanotechnology with numerous advantages has growingly attracted the attention of many researchers. Nanoscale, Size range from approximately 1 nm to 100 nm. This definition is accompanied by two notes: Note 1: Properties that are not extrapolations from a larger size will typically, but not exclusively, be exhibited in this size range. For such properties the size limits are considered approximate. Note 2:

The lower limit in this definition (approximately 1 nm) is introduced to avoid single and small groups of atoms from being designated as nano-objects or elements of nanostructures, which might be implied by the absence of a lower limit (ASTM 2006).

Nanoparticles and nanocomposites prepared chemically by precipitation or in situ formation in a given matrix through the sol-gel processes (Sanchez *et al.*, 1996). The conditions determinants of nanoparticle growth were changed in the dependence on method preparation of nanoparticles.

Materials scientists and engineers have made significant developments in the improvement of methods of synthesis of nanomaterial solids (Hansany *et al.*, 2012).

Aspergillus flavus is one of the most important pathogenic fungi that contaminate grains or feed mixtures and cause serious problems in such as mycotoxicosis due to its aflatoxins production (Hassan *et al.*, 2014). Development of new and effective antimicrobial agents seems to be of paramount importance, through the antimicrobial activity of metals having various properties, potencies and spectra of activity, has been known and applied for centuries especially in nano-size (Malarkodi *et al.*, 2014). The aim of this study is the founding of a new effective antifungal compounds through the nanotechnology against *Aspergillus flavus* which is the most dangerous fungi on animal productions.

2. Materials and methods

2.1. Synthesis of nanoparticles:

Fe₂O₃ synthesized by co-precipitation technique method by Using titra-hydrate ferrous chloride FeCl₂.4H₂O (> 99:9%) dissolved in distilled water. The ammonium hydroxide NH₄OH were added drop-wise to the mixture with stirring under strong ultrasonic agitation then added to the solution to adjust the pH value at 9 or 10 till precipitation occurred. Dark brown precipitate is formed and washed using distilled water for several times to remove excess ammonia (10 times) the precipitate was dried at 400°C for 4 hours (Kandpal *et al.*, 2014). Synthesis of Fe₃O₄ and Fe₃O₄ nanoparticles were synthesized by co-precipitation technique method of titra-hydrate ferrous chloride FeCl₂.4H₂O (> 99:9%) , about 1.9881g, and hexa-hydrate ferric chloride FeCl₃ .6H₂O (> 97:9%) , about 5.406gm (Ozkaya *et al.*, 2009).

2.2. Characterization of synthesized nanoparticles:

X-ray diffraction analyses were carried out to identify the previously prepared nanoparticles in pure single phase. Also it confirmed the successful formation of these nanoparticles according to (Cullity *et al.*, 2001)

Infrared Spectra were recorded on a Perkin – Elmer (FT-IR):

FT-IR were carried out to identify the synthesized nanoparticles and to determine its size according to (Guan *et al.*, 2003).

Evaluation of Fe₂O₃ and Fe₃O₄ nanoparticles effect on the growth of *A. flavus*.

2.3. Selection of test pathogen:

Pathogenic *Aspergillus flavus* (KP137700) selected for the study isolated from broiler feed was obtained from the Mycology Department of the Animal Health Research Institute, Giza, Egypt. Preparation of dilutions of synthesized compounds 10 mg of the each nanoparticle (Fe₂O₃ and Fe₃O₄) were weighed accurately and dissolved in 10 ml dimethyl sulfoxide giving a solution of 1mg/ml concentration. 1 ml of the above solution was again diluted to 10 ml with dimethyl sulfoxide giving a solution of 100 µg/ml concentration. Determination of minimum inhibitory concentration and fungicidal concentration Minimum inhibitory concentration (MIC) was determined for Fe₂O₃ and Fe₃O₄ nanoparticles showing antifungal activity against test pathogen by serial dilution method. Broth microdilution method was followed for determination of MIC values. Sterilized loop wire was used to transfer *A. flavus* to sabouraud dextrose agar and incubated at 25 °C for 5 days. From the *A. flavus* strain, small portion was transferred to 3ml of sabouraud dextrose broth media separately and incubated at 25°C for 24 hrs. 0.1 ml of the above five medias were transferred to five different stoppered conical flasks containing 0.9% NaCl solution. 1ml of media was taken in a test tube, to which, 1ml

of test solution (100 µg/ml) was added. Thereafter, 0.1ml of the microbial strain (*A. flavus*) prepared in 0.9% NaCl was added to the test tube containing media and test solution. Serial dilution were done five times giving concentrations of 50, 25, 12.5, 6.25, 3.75, 1.5 µg/ml. The test tube were stoppered with cotton and incubated at 25°C for one week. The MIC values were taken as the lowest concentration of the particles in the test tube that showed no turbidity after incubation. The turbidity of the contents in the test tube was interpreted as visible growth of microorganisms.

2.4. Minimum fungicidal concentration:

The minimum fungicidal concentration (MFC) was determined by sub culturing 50µl from each test tube showing no apparent growth. Least concentration of test substance showing no visible growth on sub culturing was taken as MFC.

2.5. Scanning electron microscope examination of *A. flavus* after the exposure to Fe₂O₃ and Fe₃O₄:

The treated fungi mycelia sections were collected, fixed with formaldehyde, washed with phosphate buffer solution and dehydrated with alcohol solution (30, 60, 80, 90 and 100%, maintaining the mycelia at 100%) and then submitted to critical point drying according to (Al-othman *et al.*, 2014). *Aspergillus flavus* became ready for scanning

electron microscopy (SEM) using JEOL (JSM-6380 LA) instrument.

3. RESULTS

Phase identification and structural analysis were carried out by XRD and FTIR spectra at room temperature for Fe₂O₃ and Fe₃O₄ nanoparticles showed that the investigated sample crystallized in a single phase 45nm size of Fe₂O₃ nanoparticles and 9nm size of Fe₃O₄ nanoparticles as shown in Fig 1, 2, 3 and 4.

Minimum inhibitory concentration (MIC) of Fe₂O₃, and Fe₃O₄ nanoparticles was 25mg and 50mg respectively against *Aspergillus flavus*.

SEM examination confirmed the anti-fungal effect of Fe₂O₃ and Fe₃O₄ nanoparticles through different degree of fungi cell membrane rupture fungal spores were observed damage such as reduce in spores number, malformations and hypertrophy these effects lead to destroyed and damaged of spores resulting in possible reduction of multiplication and the enzymatic activity of the micro-organism. This aspect is great important because *Aspergillus* reproduction involves mainly formation of spores. The severity of damage and malformation of *Aspergillus flavus* is high with Fe₂O₃ nanoparticles and low with Fe₃O₄ nanoparticles as in Fig 5 and 6.

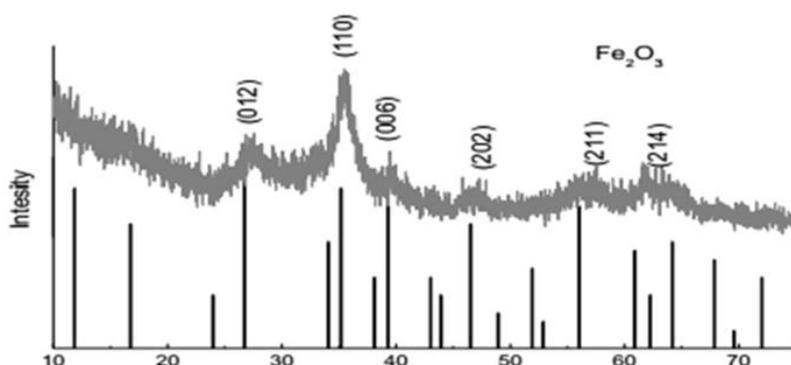


Fig.1. XRD pattern of Fe₂O₃ nanocrystalline sample

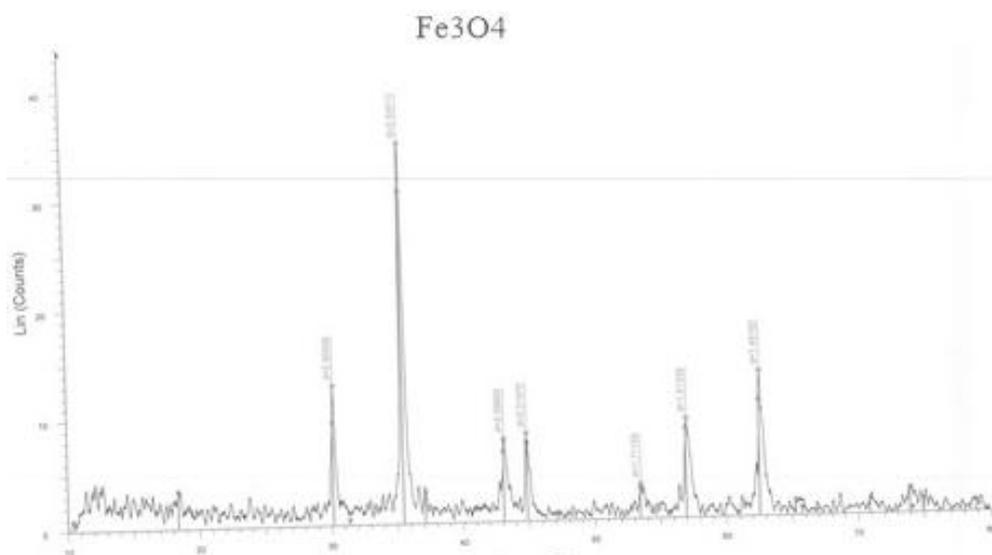


Fig.2. XRD pattern of *Fe₃O₄* monocrystalline.

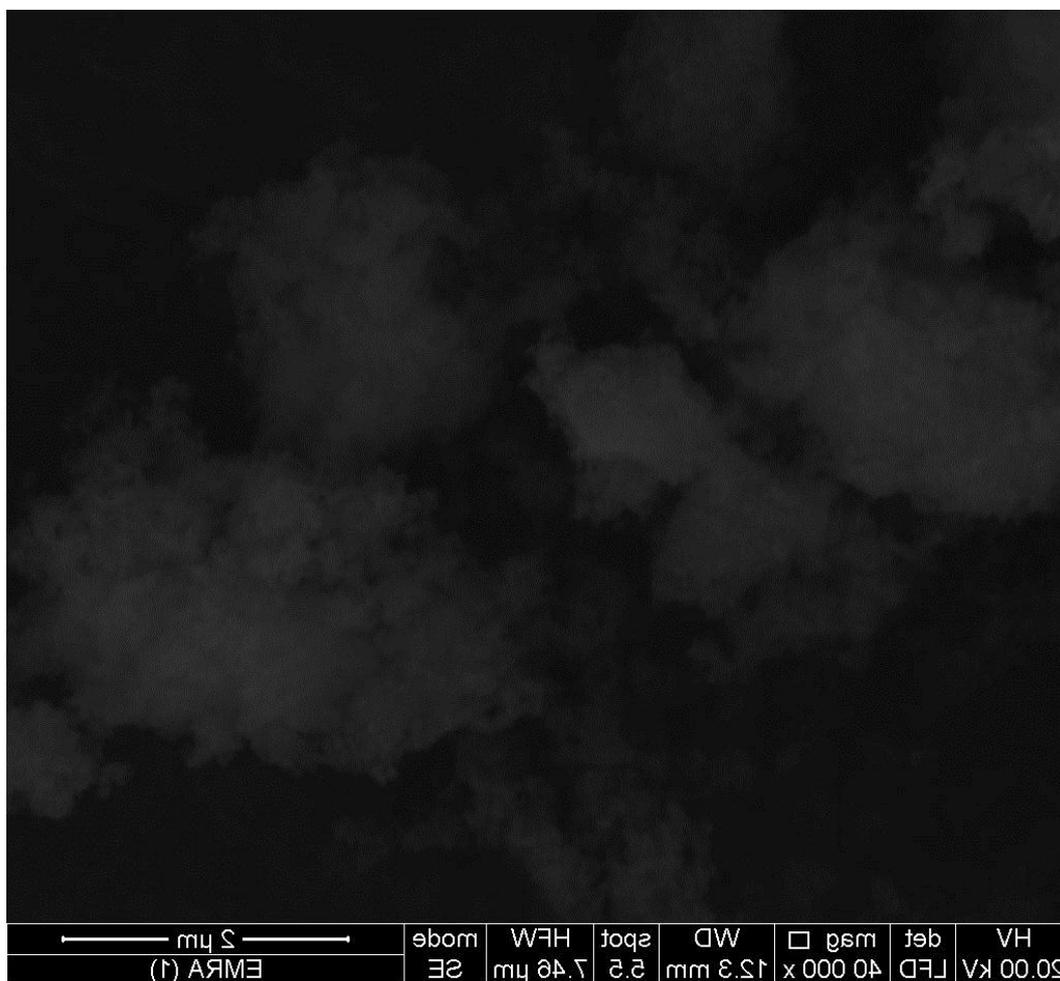


Fig.3. FTIR pattern of *Fe₂O₃* monocrystalline.

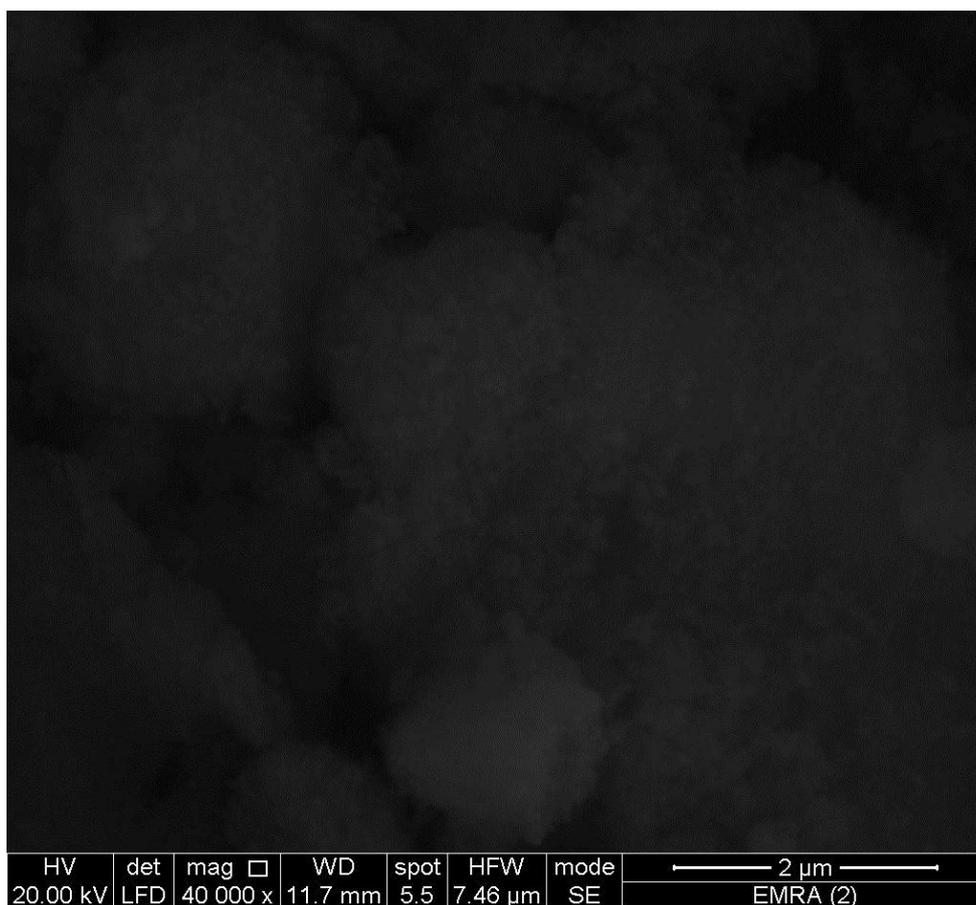


Fig.4. FTIR pattern of Fe₃O₄ monocrystalline

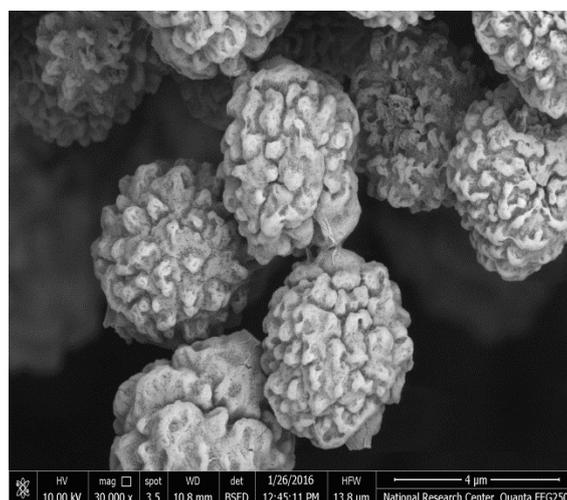


Fig.5. SEM *Aspergillus flavus* with Fe₃O₄ monocrystalline.

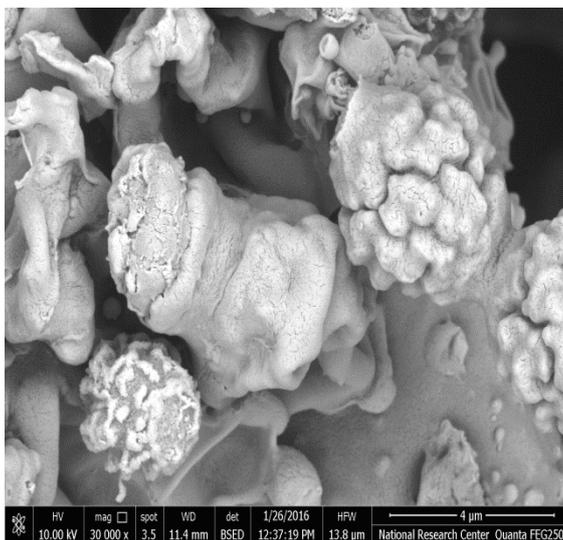


Fig.6. SEM *Aspergillus flavus* with Fe₂O₃ monocrySTALLINE.

4. DISCUSSION

According to the results of this study, it appears clearly that Fe₂O₃ and Fe₃O₄ nanoparticles have antifungal effect on *Aspergillus flavus* which is more effective in Fe₂O₃ nanoparticles and Fe₃O₄ nanoparticles.

The results of this study supported by evaluation of the antifungal potential Fe₂O₃ nanoparticles in inhibiting the growth of *Aspergillus flavus*. There was antifungal potential of prepared Fe₂O₃ nanoparticles in disc diffusion test (Gehan *et al.*, 2014). When the treated fungi with Fe₂O₃ nanoparticles were subjected to SEM, the damage and rupture of their cell wall were detected in the area surrounding growth. The normal conidial cell of *Aspergillus flavus* has a spherical shape and smooth cell wall and intact cell membrane. The effect of high concentration of Fe₂O₃ nanoparticles on the treated fungi was observed as membrane damage of cells and some pits that have been caused in intercellular components, leading to leakage and finally cell death.

The antifungal effect of Fe₃O₄ nanoparticles that evaluated in our study is agreed with (Bhupendra *et al.*, 2013) who

studied the minimum inhibitory concentration (MIC) values of Fe₃O₄-Ag nanoparticles is less than Ag nanoparticles against *Aspergillus glaucus*. The fungal growth declines as the concentration of Fe₃O₄-Ag is raised and when the latter reaches MIC, no growth is observed. Presence of Fe₃O₄ in composite with Ag gives the Ag more antifungal effect.

In conclusion, the current study shows that Fe₂O₃ nanoparticles have a strong effect on *Aspergillus flavus* as antifungal if compared with Fe₃O₄ nanoparticles which have a little antifungal effect on *Aspergillus flavus*

5. REFERENCES

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