



Prevalence of Escherichia Coli in Fish Obtained from Retail Fish Markets in Gharbia Governorate, Egypt

Saad, M.S.¹, Hassan, M.A.¹, Hassnien, F.S.¹, Abdel-Aal, M.M.², Zakar, A.H.² and Elshfey, S.A.²

¹Food Hygiene and Control Department, Faculty of Veterinary Medicine - Benha University Egypt.

²Central Laboratory for Aquaculture Research, Agriculture Research Center, Egypt.

ABSTRACT

A total of 100 random samples of fresh and marine fishes represented by *Tilapia niloticus*, *Mugil cephalus*, Sardine and Barboni (25 of each) were collected from the different fish markets located in Gharbia governorate. *E. coli* was identified by standard microbiological, biochemical tests, and further confirmed by multiplex PCR. The faecal coliform loads in fish samples were assessed. The bacterium was detected in 8(32%), 6(24%), 4(16%) and 3(12%) of the examined samples of *Oreochromis niloticus*, *Mugil cephalus*, Sardine and Barboni respectively. The accepted fish samples were 17 (68 %), 19 (76 %), 21 (86 %) and 22 (88 %) of the examined samples of *O. niloticus*, *Mugil cephalus*, Sardine and Barboni respectively while the unaccepted samples were 8 (32 %), 6 (24 %), 4 (16 %) and 3 (12 %) of the examined samples of *O. niloticus*, *Mugil cephalus*, Sardine and Barboni respectively according to Egyptian standards (E.S 3494/2005). The results cleared that, the main isolates include O26: H11; O44: H18, O86: H; O91: H21; O103: H2; O119: H6; O121: H7; O124: H; O128: H2; O146: H21 and it isolated mainly from *O. niloticus*, *Mugil cephalus*, then Sardine and Barboni. The work was aimed to study prevalence of *Escherichia coli* contamination in fish sold in different fish markets located in Gharbia governorate.

Keywords: *Escherichia coli*, *Oreochromis niloticus*, *Mugil cephalus*, Sardine, Barboni.

(<http://www.bvmj.bu.edu.eg>)

(BVMJ-34(1): 254-260, 2018)

1. INTRODUCTION:

Fish is a food of excellent nutritional value, providing high-quality protein rich in essential amino acids, and a wide variety of minerals, including phosphorus, magnesium, iron, zinc, and iodine in marine fish (Ariño *et al.*, 2013). Fish are generally considered as major vehicles for several bacterial disease transmissions. *Escherichia coli* has been generally recognized as an indicator

organism for faecal contamination of water and seafood (Geldrich 1997).

The poor unhygienic conditions of the landing centers, storage and domestic retail markets exacerbate the problem of poor hygiene and consumer safety of fish (Kumar *et al.*, 2005). Most strains of *E. coli* or faecal coliform are harmless, but some may cause diarrhea. Reed (1994) classified *E. coli* (EEC) strains to several major subgroups:

Enterohemorrhagic *E. coli* (EHEC) causes hemorrhagic colitis and hemolytic uremic syndrome. Six verotoxins have been identified within this group, but only stx-1 and stx-2 seem to be important in human infections. *E. coli* O157:H7 is the principle serotype of this group. Enteroinvasive *E. coli* (EIEC) causes a diarrheal illness similar to shigellosis. Enterotoxigenic *E. coli* (ETEC) is a major cause of travelers' diarrhea and infant diarrhea in developing countries. These strains produce a heat-labile toxin (LT) and/or a heat-stable toxin (ST). Enteropathogenic *E. coli* (EPEC) is an important cause of infant diarrhea.

The objectives of our study were to assess the prevalence of *E. coli* in fish sold at retail fish markets in Gharbia government and to enumerate the faecal coliform loads in fish.

2. MATERIALS AND METHODS:

2.1. Collection of samples:

A total of 100 random samples of Nile and marine fishes represented by *Tilapia niloticus*, *Mugil cephalus*, Sardine and Barboni (25 of each) were collected from the different fish markets located in Gharbia governorate. Each sample was kept in a separated sterile plastic bag and preserved in an ice box then transferred to the laboratory under complete aseptic conditions without undue delay and examined as quickly as possible. The collected samples were subjected to the bacteriological examination to evaluate their quality.

Preparation of samples (ICMSF, 1996):

To 5 grams of the sample, 45 ml of sterile peptone water 0.1% were added and thoroughly mixed using sterile blender for 1 - 1.5 minutes, from which tenth fold serial dilutions were prepared. The prepared samples were subjected to the following examinations

Isolation and identification of Enteropathogenic *Escherichia coli* (ICMSF, 1996). Serodiagnosis of *E. coli* (Kok et al. 1996).

Statistical Analysis:

The evaluation and interpretation of obtained results were carried out using of Analysis of Variance (ANOVA) test according to *Feldman et al. (2003)*.

3. RESULTS:

Our results cleared in table (1) indicated that, there is a significant difference of the incidences of Enteropathogenic *E. coli* isolates among examined fish samples at ($P < 0.01$).

The serotypes of *E. coli* organisms were O26: H11 in 2 (8%), 1 (4%), 1 (4%) and 0 (0%). O44:H18 1 (4%), 0 (0%), 0 (0%) and 1 (4%). O86: H 0(0%), 0(0%), 1 (4%) and 0 (0%). O91:H21 0 (0%), 1 (4%), 0 (0%) and 1 (4%). O103: H2 1(4%), 0 (0%), 0(0%) and 0 (0%). O119: H6 0 (0%), 2 (8%), 0 (0%) and 0(0%). O121: H7 1(4%), 0 (0%), 2 (8%) and 0(0%). O124: H 1 (4%), 0 (0%), 0 (0%) and 0 (0%). O128: H2 2 (8%), 1 (4%), 0 (0%) and 1 (4%). O146: H21 0 (0%), 1 (4%), 0 (0%) and 0 (0%) of the examined samples of *O. niloticus*, *Mugil cephalus*, Sardine and Barboni respectively.

Our results cleared in table (2) indicated that, there is a significant difference ($P < 0.01$) of the acceptance of fish samples based on contamination with *E. coli*.

The accepted *O. niloticus* fish were 17 (68%) and unaccepted samples 8 (32%). The *Mugil cephalus* accepted samples were 19 (76%), while the unaccepted samples were 6 (24%). While, the Sardine accepted samples were 21 (86%), while, the unaccepted samples were 4 (16%) and in Barboni the accepted samples were 22 (88%) and the unaccepted samples were 3 (12%).

Our results on Table (3), indicated that, the incidence of virulence genes of Enteropathogenic *E. coli* isolated from the examined samples of fresh and marine fish cleared that, the Shiga-toxin 1 gene (*stx1*), observed in the *E. coli* isolates of O26: H11, O44: H18, O91: H21, O103: H2, O119: H6 and O128: H2.

While, the Shiga-toxin 2 gene (*stx2*), observed in the *E. coli* isolates of O26: H11, O86, O91: H21, O103: H2, O119: H6 and

O121: H7 and O146: H21. While, the Intimin gene (*eaeA*), observed only in *E. coli* isolates of O26: H11.

The results observed in Photograph (1), indicated that, the unaccepted samples are +ve for the *E. coli* isolates as the agrose gel electrophoresis of multiple PCR gave +ve results of *stx1* (614 bp), *stx2* (779 bp) and *eaeA* (890 bp) genes for characterization of Enteropathogenic *E. coli*.

Table (1): Incidence of Enteropathogenic *E. coli* isolated from examined samples of fresh and marine fish (n=25).

Fish Spp.	<i>Oreochromis niloticus</i>		<i>Mugil cephalus</i>		Sardine		Barboni		Strain characteristics
	No.	%	No.	%	No.	%	No.	%	
<i>E. coli</i> strains									
O26: H11	2	8	1	4	1	4	0	0	EHEC
O44: H18	1	4	0	0	0	0	1	4	EPEC
O86	0	0	0	0	1	4	0	0	EPEC
O91: H21	0	0	1	4	0	0	1	4	EHEC
O103: H2	1	4	0	0	0	0	0	0	EHEC
O119: H6	0	0	2	8	0	0	0	0	EPEC
O121: H7	1	4	0	0	2	8	0	0	EHEC
O124	1	4	0	0	0	0	0	0	EIEC
O128: H2	2	8	1	4	0	0	1	4	ETEC
O146: H21	0	0	1	4	0	0	0	0	EPEC
Total	8	32	6	24	4	16	3	12	

Chi2 = 22.455**

** = Significant at (P < 0.01).

N.B. The isolation % was calculated according to the number of examined samples

EPEC=Enteropathogenic *E. coli*

EIEC =Enteroinvasive *E. coli*

ETEC=Enterotoxigenic *E. coli*

EHEC =Enterohaemorrhagic *E. coli*

Table (2): Acceptability of the examined samples of fresh and marine fish based on their contamination with *E. coli* (n=25).

Fish species	<i>E. coli</i> /g*	Accepted samples		Unaccepted samples	
		No.	%	No.	%
<i>Oreochromis niloticus</i>	Free	17	68	8	32
<i>Mugilcephalus</i>	Free	19	76	6	24
Sardine	Free	21	86	4	16
Barboni	Free	22	88	3	12

Chi2 = 9.55**

** = Significant at (P < 0.01).

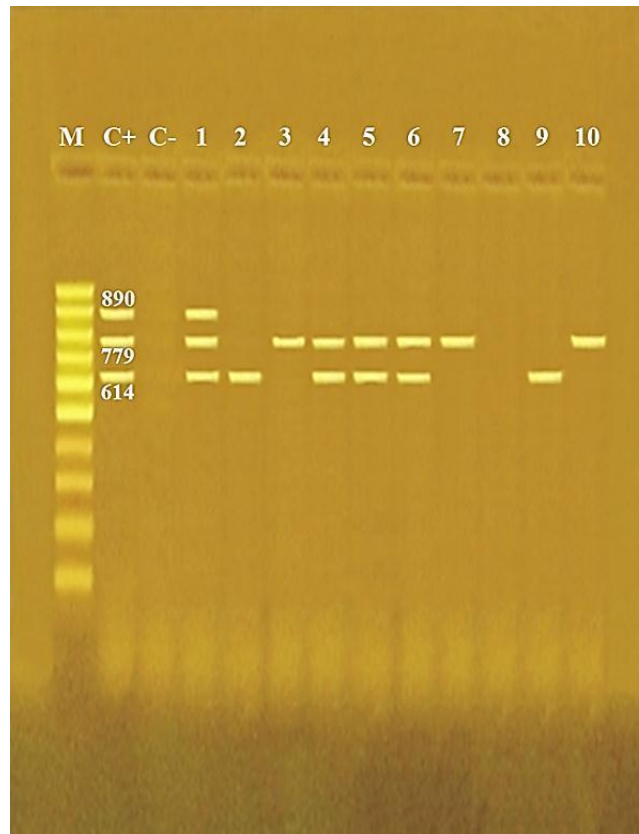
*Egyptian Organization for Standardization "EOS" (2005).

ES 3494-2005 for chilled fish

Table (3): Incidence of virulence genes of Enteropathogenic *E. coli* isolated from the examined samples of fresh and marine fish.

<i>E. coli</i> Serovars	<i>stx1</i>	<i>stx2</i>	<i>eaeA</i>
O26: H11	+	+	+
O44: H18	+	-	-
O86	-	+	-
O91: H21	+	+	-
O103: H2	+	+	-
O119: H6	+	+	-
O121: H7	-	+	-
O124	-	-	-
O128: H2	+	-	-
O146: H21	-	+	-

Stx1: Shiga- toxin 1 gene*Stx2*: Shiga- toxin 2 gene*EaeA*: intimin gene



Photograph (1): Agarose gel electrophoresis of multiplex PCR Of *stx1* (614 bp), *stx2* (779 bp) and *eaeA* (890 bp) genes for characterization of *Enteropathogenic E. coli*.

Lane M: 100 bp ladder as molecular size DNA marker.

Lane C+: Control positive *E. coli* for *stx1*, *stx2* and *eaeA* genes.

Lane C-: Control negative *E. coli* for *stx1*, *stx2* and *eaeA* genes.

Lane 1 (O26): Positive *E. coli* strain for *stx1*, *stx2* and *eaeA* genes.

Lanes 2 (O44) & 9 (O128): Positive *E. coli* strains for *stx1* gene.

Lanes 3 (O86), 7 (O121) & 10 (O146): Positive *E. coli* for *stx2* gene.

Lanes 4 (O91), 5 (O103) & 6 (O119): Positive *E. coli* for *stx1* and *stx2* gene.

Lane 8 (O124): Negative *E. coli* strain for *stx1*, *stx2* and *eaeA* genes.

4. DISCUSSION:

Quality of sea foods depends on the quality of waters from where the fishes are captured and the sanitary conditions of the landing centers. Proper Sanitation facilities at the retail markets play an important role in the overall quality of the fish. Even if the seafood samples collected from fish catch is landed in prime condition, contamination at poor landing sites and cross contamination may cause faecal contamination (Sugumar 2002). However, *E. coli* does not thrive in the marine

environment for long period of time and so this organism cannot be expected to harbor in fish harvested from the sea. The detection of *E. coli* in some fishes might represent post-harvest cross contamination (Martinez-manzaar 1992).

Our results in table (1) agreed with those of (Samaha and Hendawy, 2017), where they observed that, the most important isolates of *E. coli* from imported fish were Serotyping of *E. coli* isolated from the examined samples of imported fishes stated 5 strains of *E. coli* isolated from Basa,

Barboni and Mackerel, as O86:K61 (B7), O111:K58 (B9), O124:K72 (B17), O26:K60(B6), and O128:K67(B12). Furthermore, 4 strains were serologically isolated from Denise as O86:K61 (B7), O124:K72 (B17), O26:K60 (B6), and O128:K67 (B12).

Also, our results agreed with those of (Edris et al., 2014), where they observed that the higher bacterial count (*E. coli*) was found in Basa and gently went down in Mackerel, Barboni and Denise, respectively. Moreover, results show that the higher bacterial count (*Shigella*) was found in Mackerel and gently went down in, Basa, Barboni and Denise, respectively.

The current result of isolation of *E. coli* from the examined samples of imported fish was higher than those obtained by Singh & Kulshrestha (1994), who could isolate 17 strains from all examined samples.

As well as Ahmed (2006) who examined 15 samples of imported fish and stated that it is accepted for (E. S889/2009). Serotyping of enteropathogenic *E. coli* isolated from the examined samples of imported fish.

5. REFERENCES:

Ahmed, A. 2006: Improvement of The Bacteriological Quality of Fish Fillets and Peeled Prawn Available in The Egyptian Markets, PhD thesis, fac Vet Med, Cairo University, Egypt.

Ariño, A., Beltrán, J.A., Herrera, A., Roncalés, P. 2013: Fish and seafood: Nutritional Value-Encyclopedia of Human Nutrition (Third Edition), Pp: 254-261.

Edris AA, Reham A, Amin Marionette Z, NaseifEbtsam M, Abdel Fatah 2014 Evaluation of Retailed Salted Fish according to Egyptian Standard.

Benha Vet Medical J 27 (2): 168-176.

Feldman, D.; Ganon, J.; Haffman, R. and Simpson, J. 2003: The solution for data analysis and presentation graphics. 2nd Ed., Abacus Lancripts, Inc., Berkeley, USA.

Geldrich, E.E. 1997: Coliforms: a new beginning to an old problem. In Coliforms and *E. coli*: Problem or Solution, eds. Kay, D. & Fricker, Cpp. 3–11.

International commission of Microbiological Specification for Foods "ICMSF" 1996: Microorganisms in Food. I. Their Significance and methods of enumeration. 3rd Ed. Univ. of Toronto, Canada.

Kok, T.; Worswich, D. and Gowans, E. 1996: Some serological techniques for microbial and viral infections. In Practical Medical Microbiology (Collee, J.; Fraser, A.; Marmion, B. and Simmons, A., eds.), 14th ed., Edinburgh, Churchill Livingstone, UK.

Kumar H. S., Parvathi A., Karunasagar I., and Karunasagar I. 2005: Prevalence and antibiotic resistance in *Escherichia coli* in tropical Seafood. World Journal of Microbiology and Biotechnology. 21 pp 619-623.

Martinez-manzaares, E. Moringo, M.A. Castro, D. Balebona, M.C. Munoz M.A. and Borrego, J.J. 1992: Relationship between indicators of faecal pollution in shell fish, growing water and the occurrence of human pathogenic microorganisms in shell fish. Journal of Food Protection., 55 pp 609-614.

Reed, G.H. 1994. Foodborne illness (part 8): *Escherichia coli*. Dairy, Food and Environmental San. 14(6):329-330.

- Samaha IA and Hendawy TA. 2017: Enterobacteriaceae in Some Imported Fish. Dairy and Vet Sci J. 2017; 3(5): 555624.
- Singh BR, Kulshrestha SB 1994: Prevalence of *Shigelladysentriae* group A type in freshwater fishes and seafood. J Food Sci Technol 30 (1): 52-53.
- Sugumar, A. 2002: Sanitary status of fish landing sites and microbial quality of fresh fish of commerce; suggestion for improvement, SDMRI Research Publication., 2 pp 153-158.