



Pathological and biochemical studies on ochratoxicosis in balady duckling with trail of treatment

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ABSTRACT

A total of 100 healthy balady ducklings, one day old were divided into 4 equal groups (25 in each), Gp (1) kept as control group, Gp (2 and 3) received 0.5 mg ochratoxin/kg ration and 0.5ml antiochratoxin/liter drinking water for 30 consecutive days respectively and Gp (4) received both ochratoxin and antiochratoxin in same dose, period and route of administration. Ducklings supplemented with ochratoxin displayed significant reduction in weight gain, feed conversion rate, RBCs, WBCs, Hb, PCV, T protein albumin, globulin, T. lipid, cholesterol, triglycerides CAT and SOD, coupled with significant elevatoin in feed conversion rate, mortality rate, AST, ALT, ALP, uric acid, creatinine and MDA Ducklings received anti-ochratoxin showed significant increase in weight gain, RBCs, Hb, total protein, albumin Catalase and super oxide dismutase beside in signif- icant increase in PCV,WBCs, globulin, AST, ALT, ALP, uric acid and creatinine, triglycerides cholesterol and total lipid coupled with insignificant decrease in MDA and improved in feed conversion rate. Hemato-biochemical profiles and antioxidant revealed significant improvement in antiochratoxin treated ducklings when compared with ochratoxinated duckings. Ochratoxin residues in liver and kidney were high at 1st day of clearance period and disappeared at 10th days of clearance period. Alteration in histological picture in internal organ in duckling received ochratoxin revealed massive degenerative changes in renal epithelial cells of proximal and distal convoluted tubules (Nephrotoxicity) and massive necrosis of some tubular epithelial cells. Beside severe hepatic damage and also lesions in the immune organs and intestine. In general, histological changes were mild in birds receiving antiochratoxin alone or with ochratoxin. It could be concluded that, ochratoxicosis in duckling resulted in poor feed intake, weight gain and feed conversion rate, had adverse effect in body weight, hemato-biochemical parameters beside decreased blood antioxidant; anti-ochratoxin improved these parameters in duck.

Keywords: duckling, Ochratoxin, body weight, mortality rate, blood picture, biochemical parameters, antioxidant enzymes, Pathology.

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1. INTRODUCTION

Mycotoxins are often found as natural contaminants in grains as secondary metabolites of fungi with major threat to livestock and human beings (Sinha, 1972). The mycotoxins include a complex group of chemical substances causing problems in poultry farms (Schwrdt et al., 2007). The family of ochratoxins consists of three members known as ochratoxin A, B and C but ochratoxin A is the most toxic one causing many adverse effects in domestic livestock causing subacute and chronic production losses in poultry (Huff et al., 2006). Ochratoxin interfere with synthesis of enzymes and other proteins by competitively inhibiting phenylalanine-tRNA (Pattison et al., 2008). Ochratoxins are nephrotoxic, carcinogens and immunotoxins in rats and human (Zahoor et al., 2012). Ochratoxin reduce growth rate, immune responses and

increase the mortality rate (Tansakul et al., 2012). Decreased humoral immune response to suppression of phagocytosis and poor response to vaccines are common manifestations of ochratoxicosis (Girish and Smith, 2008). The present study was done to evaluate the toxic effects of ochratoxin alone on body performance, hematolo-biochemical and pathological changes in balady duckling and modulation this toxic effect using antiochratoxin.

2. MATERIALS AND METHODS

2.1. Anti-ochratoxin

It is produced under trade name Texal from Miles Company (Aspania) iumported by Optima Compay, 6th october, Egypt, compound consisted of pure oligosaccharide, lactobacillus, 'actic acid and yeast.

Ochratoxin A was obtained from microbiology Dep. Fac. of Vet. Med. Zag. Univ.

2.2. Ducklings and Experimental design

One hundred balady duckling, one day old, average weight 50 gm were housed under hygienic condition, fed on freshly formulated balanced ration free from any mycotoxin residues and given water ad-libitum. Duckling were divided into 4 groups (25 ducks each), Gp (1) (control group), Gp (2) received 0.5 mg ochratoxin /kg ration, Gp (3) received 0.5ml antiochratoxin /liter drinking water for 30 consecutive days and Gp (4) received ochratoxin (0.5mg/kg ration) plus antiochratoxin (0.5ml antiochratoxin/liter) in same period and rout. Supplementation of ochratoxin and antiochratoxin for 30 days (from 1st day of age up to 30th day of age). Mortality rate was recoded in all groups from start of experiment to 30 day of age. Body weight: Five ducklings in each group were weighted individually at start of the experiment (1stday of age) & at1st day post Supplementation (31thday of age), consumed ration was recorded for calculation of weight gain and feed conversion rate (FCR)

2.3. Sampling

At 1st day post supplementation (31th day of age) five ducklings from each group were slaughtered and two blood samples were collected. 1st sample was taken in tube contain EDTA as anticoagulant for estimation estimation (RBCs, Hb, PCV% and TWCs) (Jain, 2000), catalase (CAT) (Solcan, et al. 2008) superoxidase dismutase (SOD) (Palabiyik, et al. 2013) Malonodialdehyde (MDA) (Esterbauer, 1982). The 2nd sample was taken to obtain clear serum for estimation of total protein (Dumas, et al. 1981), albumin (Drupt, 1974), globulin was calculated as difference between total protein and albumin, total lipid (Knight, et al.1972), triglyceride (Royer, 1969), cholesterol [43], transaminases (AST and ALT) (Reitman and Frankel 1957), ALP (John, 1982), Uric acid (Coalombe and Faurean, 1963) and creatinine (Husdan and Rapoport, 1968).

2.4. Ochratoxin residue:

At 1st, 5, 10th and 15th day post ochratoxin, 3 ducklings from each group were slaughtered and sample from liver and kidneys were taken for determination of ochratoxin residues (Jürgensen, 2004).

2.5. Pathological examination

Specimens were taken from liver, kidneys, intestine, spleen and bursa of the sacrificed duckling and directly fixed in 10% neutral buffered formalin. Five-micron thick paraffin sections were prepared stained with hematoxylin and eosin and examined

microscopically (Bancroft and Gamble, 2002).

2.6. Statistical analysis

The obtained data was analyzed according to Petrie and Watson (1999).

3. RESULTS

Mortality rate Body performance, hemato-biochemical parameters and ochratoxin residues were recorded in tables (1-5). Grossly, liver and kidney OF duck received ochratoxin (Gp 2) were enlarged, cyanosed and some cases pale in color. The gall balder enlarged and filled by bile macroscopically change in Gp 2 were sever, the kidney in showed coagulative necrosis of some renal tubule, severe congestion and hemorrhage (Fig, 1). Kidney showed coagulative necrosis of the renal tubules accompanied by lymphocytic aggregations (Fig, 2). Kidney showed severe hydropic degenerative changes in the renal tubules, beside vacular and hydropic degeneration (Fig, 3). Kidney showing perivascular edema and nephritis (Fig, 4). The hepatic tissue showed toxic hepatitis manifested by extravasated erythrocytes, leukocytic infiltration and vacular degeneration accompanied by interstitial fibrosis (Fig, 5). Bursa of fabricius showed lymphocytic depletion and hyperplasia of the mucosal lining epithelium (Fig, 6). Spleen showed necrosis, perivascular edema and lymphoid depletion (Fig, 7), intestine showed epithelial desquamation and necrosis (Fig, 8), kidney showed congestion, vacular and hydropic degeneration as well as lymphocytic infiltration (Fig, 9). The histopathological changes that noticed in the group 4 become mild or less severe the bile ductule showed proliferation portal to portal lymphocytic infiltration (Fig, 10). Intestine showed leukocytic infiltration and periglandular fibrosis (Fig, 11). Bursa of fabricius showed nearly normal structure (Fig, 12).

4. DISCUSSION

In the present study ochratoxin resulted in significant increase in mortality rate 7 (17.50%). Ochratoxin induce about 12.98% mortality in broilers (Kumar et al., 2003). Meanwhile it has been found ochratoxin caused significant increase in mortality up to 40% in turkey poults (Chang et al., 1981). Similar mortality rate was recorded in duckling (Yang et al., 2013).

Ochratoxin induced significant reduction in body weight gain, feed consumption and elevation in FCR in duckling. Antiochratoxin (Texal) induce significant elevation in weight gain beside improved FCR. Reduction in weight gain during ochratoxicosis may be due to adverse effect of ochratoxin in

Table (1) Effect of Ochratoxin and anti-ochratoxin (Texal) on mortality and body performance in duckling (n =5)

Groups	Total No	Mortality rate		Ineasial B.W(gm)	Weight at 1st day PT (gm)	BWG (gm)	FC (gm/duckling)	FCR
		No	%					
Gp (1)	25	00	00	51.43±0.59	205.94±0.84	154.51±7.57	199.05	1.29
Gp (2)	25	7	28	50.53±0.62	185.05±0.93	134.52±2.42*	185.14	1.37
Gp (3)	25	1	4	51.49±0.55	248.17±0.84	196.68±8.59**	218.42	1.11
Gp (4)	25	3	12	51.05±0.49	214.74±0.38	163.69±7.61	209.27	1.28

*** Significant at $P < 0.001$. PT=post treatment BWG= body weight gain FC = feed consumption FCR = feed conversion rate

Table (2) Effect of Ochratoxin and anti-ochratoxin (Texal) on blood picture, in duckling (n =5)

Parameter	Control duckling	ochratoxin	Anti- Ochratoxin	Ochratoxin+ anti
RBCs (10^6 /cm.m)	4.05 ± 0.18	3.22 ± 0.20*	5.03± 0.31*	4.25± 0.42
Hb (gm/dls)	13.16 ± 0.78	10.23 ± 0.60*	15.19 ± 0.41*	14.04 ± 0.64
P.C.V. %	36.31± 0.96	32.16± 0.84*	37.06± 0.42	36.68± 0.82
TWCs (10^3 /cmm)	10.04±0.56	8.86±0.13*	11.10±0.73	10.31±0.69

*Significant at $P \leq 0.05$

Table (3): Effect of Ochratoxin and anti-ochratoxin (Texal) on some biochemical parameters, in duckling (n =5)

Parameter		Gp (1)	Gp (2)	Gp (3)	Gp (4)
Protein profile (mg/dl)	T. protein	5.98±0.46	4.04±0.39**	7.31±0.37*	6.03±0.37
	Albumen	3.43±0.22	2.54±0.24*	4.34±0.26*	3.64±0.46
	globulin	2.55±0.25	1.50±0.34*	2.97±0.15	3.06±0.24
	A/G ratio	1.35±0.19	1.63±0.21	1.46±0.32	1.19±0.32
Liver enzymes (U/L)	AST	37.05±1.07	40.38±0.89*	36.98±0.94	37.21±0.86
	ALT	31.47±1.15	36.14±1.05*	30.42±0.52	31.65±0.91
	ALP	36.19±1.53	42.17±1.17*	36.05±0.47	36.44±0.83
Kidney Function (mg/dl)	Uric acid	4.62±0.33	5.53±0.21*	4.55±0.37	4.98±0.51
	Creatinine	1.04 ± 0.15	1.55±0.12*	1.02±0.16	1.09±0.23
Lipid profile	total lipid	632.04±2.17	625.16±1.04*	635.21±1.07	631.07±1.42
	triglyceride	98.52±1.35	92.05±1.81 *	99.70±1.73	96.42±1.53
	cholesterol	149.21±1.82	142.06±1.86*	152.32±1.27	148.72±1.50

*Significant at $P \leq 0.05$

Table (4): Effect of Ochratoxin and anti-ochratoxin (Texal) on some antioxidant enzymes, in duckling (n =5)

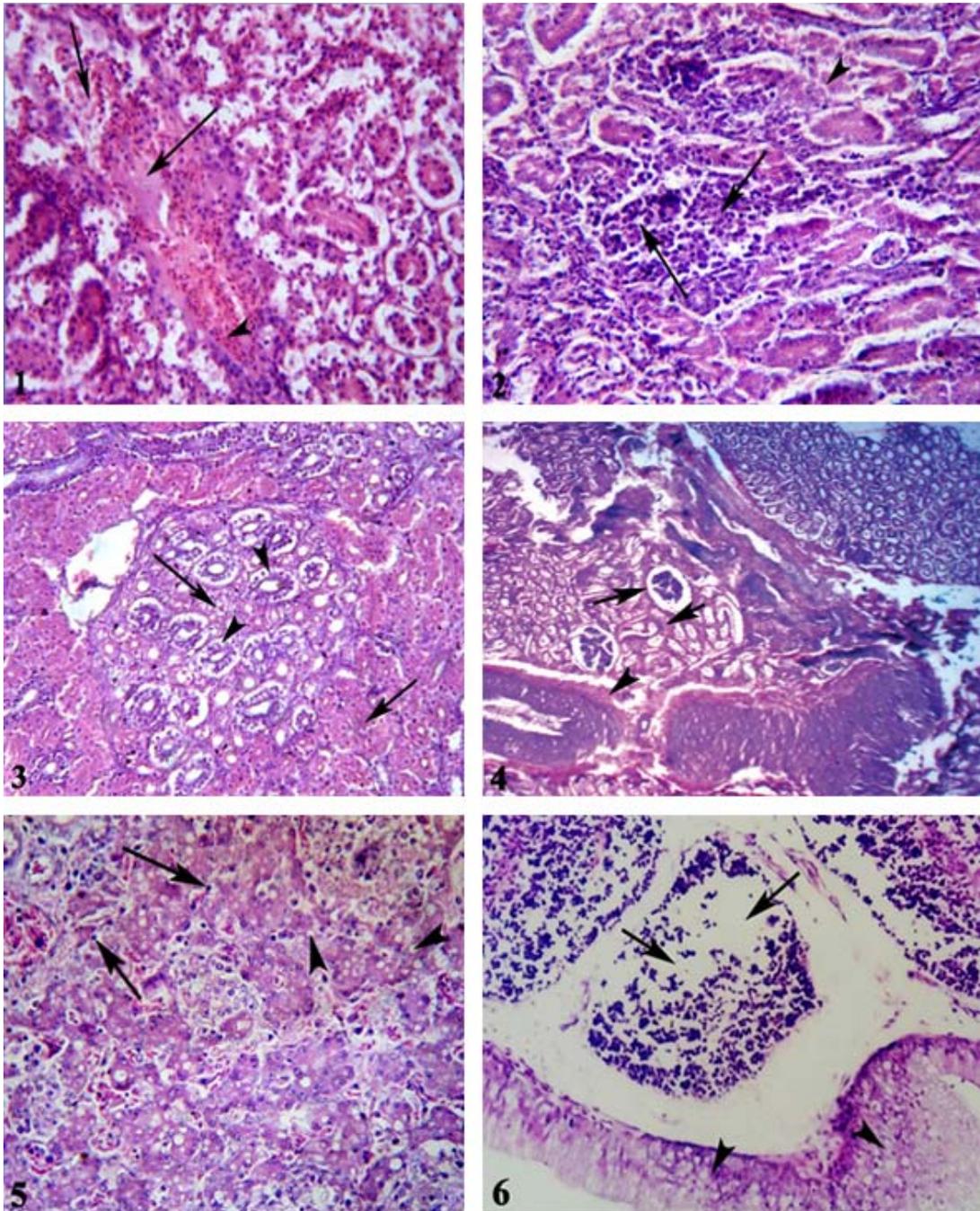
Parameter	Gp (1)	Gp (2)	Gp (3)	Gp (4)
catalase	1.83 ± 0.21	1.05 ± 0.19*	2.42 ± 0.14*	2.01 ± 0.27
Superoxid dismutase (U/mL)	70.10± 0.39	68.52±0.51*	71.71± 0.34*	70.52± 0.72
Malondialdehyde (nmol/mL)	13.57± 0.41	15.26±0.37*	13.38± 0.51	13.18± 0.62

*Significant at $P \leq 0.05$

Table 5) Ochratoxin residus in liver and Kidney (ppm/gm) during and post supplementation of duckling (n=5)

Days post treatment	Gp (1)		Gp (2)		Gp (4)	
	Liver	Kidney	Liver	Kidney	Liver	Kidney
1	00	00	4.66±0.22	5.21±0.36	3.05±0.42	3.15±0.31
5	00	00	1.41±0.16	1.63±0.21	0.83±0.31	0.90±0.15
10	00	00	ND	ND	ND	ND
15	00	00	ND	ND	ND	ND

ND= Non-detected



Fig, 1, kidney of duckling in Gp 2 showing coagulative necrosis of some renal tubule, severe congestion and hemorrhage (H&EX400). Fig, 2, kidney of duckling in Gp 2 showing coagulative necrosis of the renal tubules accompanied by lymphocytic aggregations (H&EX400). Fig, 3, kidney of duckling in Gp 2 showing severe hydropic degeneration in the renal tubules, beside vacuolar and hydropic degeneration (H&EX400). Fig, 4, kidney of duckling in Gp 2 showing perivascular edema and nephritis (H&EX200). Fig, 5, liver of duckling in Gp 2 showing toxic hepatitis manifested by extravasated erythrocytes, leukocytic infiltration and vacuolar degeneration accompanied by interstitial fibrosis (H&EX400). Fig, 6, bursa of fabricius of duckling in Gp 2 showing lymphocytic depletion and hyperplasia of the mucosal lining epithelium (H&EX600)

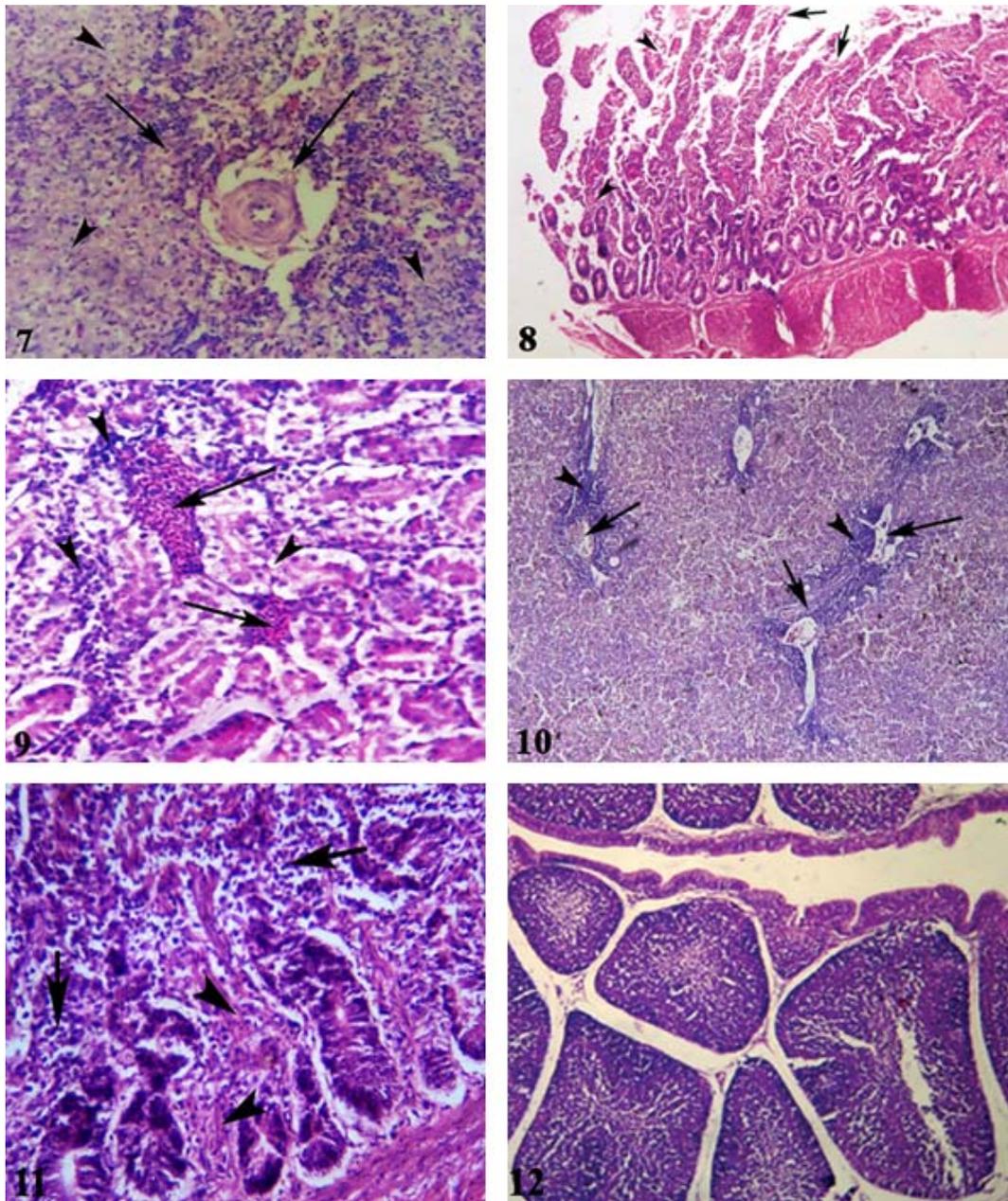


Fig. 7, spleen of duckling in Gp 2 showing necrosis, perivascular edema and lymphoid depletion (H&EX600). Fig. 8, intestine of duckling in Gp 2 showing severe epithelial desquamation and necrosis (H&EX300). Fig. 9, kidney of duckling in Gp 4 showing congestion, vacuolar and hydropic degeneration as well as lymphocytic infiltration (H&EX400). Fig. 10, liver of duckling in Gp 4 showing bile ductule proliferation portal to portal lymphocytic infiltration (H&EX200). Fig. 11, intestine of duckling in Gp 4 showing leukocytic infiltration and periglandular fibrosis (H&EX400). Fig. 12, bursa of fabricius of duckling in Gp 4 showing nearly normal structure (H&EX300).

intestinal tract decreasing feed absorption that led to decrease weight gain alterations (Raju and Devegowda, 2000). Reduction in weight gain due to ochratoxicosis was in agreement with previous reports using dietary ochratoxin of broilers (Garcia et al., 2003). Elevation in body weight gain and improved FCR due to oligosaccharide in anti-ochratoxin due to improved intestinal function or gut

health (Huff et al., 1988). Improvement in body performance was found in quail received oligosaccharide (Ghosh et al., 2007).

Ochratoxin resulted in significant decrease in Rbcs, WBCs, Hb, PCV % in duckling but Antiochratoxin (Texal) induced significant increase in RBCs and Hb beside insignificant increase in

WBCs and PCV % Reduction in RBCs and Hb content in ochratoxicosis has been noted in broiler chickens (Elaroussi et al., 2006). Reductions in RBCs and Hb content may be due to reduction in serum iron in ochratoxicosis (Agag 2004). Reduction in total circulating WBCs during ochratoxicosis, due to decreases in lymphocytes and monocytes (Chang et al., 1981). Change in blood picture post using antiochratoxin may be due to presence of oligosaccharide and lactobacillus. Similar changes in blood picture were previously recorded (Banergee et al., 2002) in chickens received oligosaccharides. Same change in erythrogram was recorded in rabbit received oligosaccharides (Bovera et al., 2010).

In the present study, duckling received ochratoxin evoked significant reduction in total protein; albumin and globulin, meanwhile antiochratoxin (Texal) induce significant increase in serum total protein, albumin and globulin. Ochratoxin feeding resulted in dose dependent decrease in total protein and albumin in white leghorn hens (Hassan et al., 2012). Reduction in protein profile during ochratoxicosis may be due to reduced feed intake (Raju and Devegowda, 2000) and/or due to decline in protein biosynthesis as ochratoxin induce inhibition hepatic protein synthesis (Prior et al., 1980). Oligosaccharide led to a significant increase in serum total protein, albumin and globulin (Shahzad et al., 2014).

In the present study, it has been shown that, ochratoxin resulted in a significant increase in the activity of liver enzymes (AST, ALT and ALP), uric acid and creatinine in duckling but antiochratoxin (Texal) induce insignificant effect in liver enzymes (AST ALT and ALP), uric acid and creatinine. Elevated liver enzyme may be due to tissue damage and leakage of enzymes into the blood stream (Sawarkar et al., 2011). Our results agreed with Sakhare et al. (2007) who reported that ochratoxin increased uric acid and creatinine in broiler chicks. Ochratoxin induce kidney damage and increased uric acid and creatinine (Raju and Devegowda, 2000).

In the present study, it has been shown that ochratoxin induced a significant decrease in serum total lipid, cholesterol and triglycerides. Our results were supported by the results achieved by Elaroussi et al. (2008) mentioned that ochratoxin induce reduction in total lipid, triglycerides and cholesterol in chickens. Ochratoxicosis induce reduction in triglycerides and cholesterol in broiler. Reduction in serum total lipid, cholesterol and triglycerides was reported in ochratoxinated broiler (Schaeffer et al., 1987). Serum total lipid, triglycerides and cholesterol levels of white Pekin ducks were not affected by dietary oligosaccharide [Al-(Al-Rawashdeh et al., 2000; Costa et al., 2015).

Our results revealed ochratoxin induce significant decrease in CAT and SOD beside significant increase in MDA but antiochratoxin (Texal) induces significant increase in CAT and SOD beside insignificant decrease in MDA. Same observation was recorded stated that ochratoxin induced decreased in CAT and SOD and increase in MDA (Dhanalakshmi et al., 2015; Soyöz et al., 2004). Our results go hand in hand with those reported [36] stated that ochratoxin induce decreases in CAT, SOD and elevation in MDA in rat. Change in antioxidant enzymes post using antiochratoxin reported (Singh and Chauhan, 2011).

The main gross pathological lesion appeared in duckling received ochratoxin were enlargement of the liver and kidney. Same gross pathological lesion was observed previously in chickens (Biro et al., 2002). Pathological finding in our work revealed severe lesion in duckling received ochratoxin but in duckling received ochratoxin with antiochratoxin lesions were mild. These lesions were represented by sever nephrogenic change manifested by degenerative change in renal tubule. Hepatic tissue appears suffering from more lesion as congestion, vacuolar and hydrobic degeneration, bile ductule proliferation and necrosis in some cases. Bursa and spleen undergo lymphoid depletion. Intestinal mucosa under go epithelial desquamation and lymphocytic infiltration. Francisco and Maria (2010) reported that ochratoxin induce acute proximal tubular epithelial necrosis in the kidneys and theses lesion could be related to oxidative damage of ochratoxin. Ochratoxin induces degenerative changes in kidney and liver (Solcan et al., 2008). Same pathological changes were observed in ochratoxicated ostrich (Elwan et al., 2009).

Ochratoxin residues in liver and kidney were high at 1st day of clearance period and disappeared at 10th days of clearance period. High residue was found in kidney then liver due to fact kidney is the main target organ of ochratoxin (Alvarez et al., 2004). The obtained results coincide with Juskiewicz et al. (1982) recorded that ochratoxin eliminated within 7 days post ochratoxin removal from the diet. Higher ochratoxin residues in kidneys than in liver (Zahoor et al., 2012).

Our study demonstrated antiochratoxin ameliorates adverse effects of ochratoxin in duckling and providing largely return the body weight gain and feed conversion rate (Agawane and Lonkar, 2004), reduction in mortality rate up to 3(7.5%), erythrogram, total leukocytic count and biochemical parameters toward the normal levels (Farag et al., 2009). Such ameliorative effect of ochratoxin could be attributed to oligosaccharide present in

antiochratoxin (Singh and Chauhan, 2011).

From this study, we concluded that, ochratoxin in duckling resulted in adverse effect in body weight, some hemato-biochemical and antioxidant enzymes parameter, antiochratoxin (Texal) treatment in duckling improved these parameters.

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