



## Biochemical Studies on Evaluation of New Composite against Hepatocellular carcinoma-induced in Rats: Recent Therapeutic Approaches

Omaima A. Ragab abou Zaid<sup>1</sup>, Yakout A. El- Senousy<sup>1</sup>, Abdel Fattah M. Badawy<sup>2</sup> and Ahmed Mohammed Rashad<sup>3,\*</sup>

<sup>1</sup> Biochemistry Department, Faculty of Vet. Medicine, Benha University, Egypt, <sup>2</sup> Applied Chemistry Department, Egyptian Petroleum Research Institute, Egypt, <sup>3</sup> Biochemistry Department, Faculty of Pharmacy, October 6 University, Egypt.

### ABSTRACT

The hepatoprotective effect of some natural and synthetic compounds against chemically induced hepatocellular carcinoma in rats was evaluated. One hundred male albino rats were divided equally into five groups. Normal control group, carcinogenic [Ferric nitrilotriacetate (Fe-NTA: 9 mg Fe/kg b.wt. i.p.) and chloroform (150 mg/ kg b.wt. orally)] - induced group, curcumin group (400 mg/kg. b.wt. orally), tetrachlorocuprate-lysine (25 mg/kg. b. wt. s.c.) and ascorbate (500 mg/kg. b. wt. orally) group and a mixture group (composed of curcumin, tetrachlorocuprate -lysine and ascorbate). Blood samples and liver tissue specimens were collected at the end of experiment (4 months) for determination of the following parameters: Tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 (IL-1), catalase and myeloperoxidase (MPO) in liver tissues, in addition to serum malondialdehyde (MDA) and reduced glutathione (GSH). Moreover, histopathological examination of liver tissues was done for results confirmation. The obtained results showed a significant elevation in MDA, MPO and immunological markers levels, with significant reduction in serum reduced glutathione and catalase activity in liver tissue in hepatocellular-carcinogen induced rats as compared to the control group. However, administration of rats with the compounds under investigation resulted in a significant reduction of MDA, MPO and immunological markers levels, and increased in reduced glutathione and catalase levels compared to the carcinogenic non treated group. Various pathological alterations were observed in liver of chemically induced-carcinogenic group interestingly, results supported the protective effect of the compounds under investigation and preserved the histological structures of liver tissues. These results concluded that basic curcumin, tetrachlorocuprate-lysine and ascorbate exert chemopreventative effect against hepatocellular carcinoma.

**Keywords:** Ferric nitrilotriacetate, curcumin, antioxidant, hepatocellular carcinogen.

(<http://www.bvmj.bu.edu.eg>)

(BVMJ-30(1): 124-136, 2016)

### 1. INTRODUCTION

Hepatocellular carcinoma (HCC), also called malignant hepatoma, is the most common type of liver cancer. Most cases of HCC are secondary to either a viral hepatitis infection (hepatitis B or C) or cirrhosis (alcoholism being the most common cause of liver cirrhosis (Kumar et al., 2003). Ferric nitrilotriacetate (Fe-NTA) is one such environmental toxicant, a known substitute for pyrophosphate used in various kinds of detergents, Fe-NTA induces lipid

peroxidation (LPO) which in turn generates oxidative stress and reactive oxygen species (ROS), also produces oxidative modifications in deoxyribonucleic acid (Iqbal et al., 1999). Fe-NTA plays an essential and fundamental role in nephrotoxicity and tumorigenesis (Kaur et al., 2007). Chloroform is an organic compound with formula  $\text{CHCl}_3$ . It is one of the four chloromethanes (Rossberg et al., 2006). Chloroform may be released to the air as a result of its formation in the

chlorination of drinking water, wastewater and swimming pools. Other sources include pulp and paper mills, hazardous waste sites, and sanitary landfills. The major effect from acute (short-term) inhalation exposure to chloroform is central nervous system depression. Also, chloroform has been shown to be carcinogenic in animals after oral exposure, resulting in an increase in kidney and liver tumors (Agency for Toxic Substances, 1997). Curcumin is the principal curcuminoid of the popular Indian curry spice turmeric, which is a member of the ginger family (Zingiberaceae). Curcumin can exist in at least two tautomeric forms, keto and enol. The enol form is more energetically stable in the solid phase and in solution. Akram et al., (2010) have suggested that in vitro and animal studies, the curcumin may have antitumor, antioxidant, antiarthritic. Curcumin inhibited the recruitment of RNA polymerase II to viral DNA, thus inhibiting the transcription of the viral DNA. Curcumin acts as a free radical scavenger and antioxidant, inhibiting lipid peroxidation and oxidative DNA damage. Curcuminoids induce glutathione S-transferase and are potent inhibitors of cytochrome P450. Ascorbic acid is a naturally occurring organic compound with antioxidant properties. It is a white solid, but impure samples can appear yellowish. It dissolves well in water to give mildly acidic solutions. Ascorbic acid is one form of vitamin C (Lachapelle et al., 2013). Tetrachlorocuprate, Cuprate loosely refers to a material that can be viewed as containing copper anions. Examples include tetrachlorocuprate ( $[\text{CuCl}_4]^{2-}$ ) (Potassium Cuprate (III)" in Handbook of Preparative Inorganic Chemistry). Copper is a component of various intracellular and extracellular enzymes such as cytochrome oxidase, lysyl oxidase, ceruloplasmin and superoxide dismutase (Klasing et al., 1998). Copper complexes are often toxic to cells, therefore tumor cells were killed, while normal cells in the whole body remained

alive for the lower level of copper (Daniel et al., 2005).

## 2. MATERIAL AND METHODS

### 2.1. Chemical and Reagents

Nitrilotriacetate was obtained from Thermo Fisher Scientific Inc- England and Ferric Nitrate from El-Nasr chemical company- Egypt.

#### 2.1.1. Preparation of Fe-NTA solution:

Fe-NTA was prepared freshly and immediately before its use. To prepare Fe-NTA, ferric nitrate solution was mixed with four-fold molar excess of disodium salt of NTA and the pH is adjusted to 7.4 with sodium bicarbonate solution (Awai et al., 1979). Chloroform was obtained from Thermo Fisher Scientific Inc- England. Ascorbic Acid was obtained from TS laboratory company- Egypt. Curcumin was obtained from Loba Chemie Company – India and mixed with Sodium bicarbonate at a concentration of (1:4) to form basic curcumin. Tetrachloro cuprate and Lysine were obtained from Sigma Aldrich-USA.

#### 2.1.2. Preparation of Bis (L-Lysine-Tetrachlorocuprate):

L-Lysine HCl and  $\text{CuCl}_2$  in a 2:1 molar ratio were mixed together and grinded in an agate mortar for about 30 minutes at room temperature to get the best homogeneity. The mixture was gradually turned to light green indicating the formation of Tetrachlorocuprate complex (Adams et al., 2010).

### 2.2. Animals and Grouping

This work was carried out on 100 male albino rats, weighting 40-60gm., purchased from the animal house colony of the National Cancer Institute (NCI), Cairo University, Egypt. Animals were housed under normal environmental conditions of standard temperature, humidity and diurnal environment of light and dark and fed a standard diet, which composed of (24% proteins, 5.55% fibers, 5.5% ash) and drink tap water ad libitum. Rats were

divided into five main equal groups, 20 rats each and classified as follows: Group (1): Rats were fed standard food and drinking tap water ad-libitum for 4 months and served as control group. Group (2): Rats were given intraperitoneal injection of Fe-NTA (9 mg Fe/kg body weight) (Summya *et al.*, 2013) and chloroform (150 mg/kg b.wt.) dissolved in corn oil, orally (Afrah *et al.*, 2014) for 4 months. Group (3): Rats were administered orally with basic curcumin (400 mg/kg) (Yumei *et al.*, 2008) half an hour prior to Fe-NTA and chloroform administration and continued till the end of experiment (4 months). Group (4): Rats were received Bis (tetrachlorocuprate- Lysine) dissolved in ethylacetate (25 mg/kg. b. wt. s.c.) (Frechilla *et al.*, 1999) and ascorbate (500 mg/kg. b. wt.) (Adejuwon *et al.*, 2008) orally and daily half an hour prior to the administration of Fe-NTA and Chloroform till the end of 4 months. Group (5): Rats treated daily with Bis (L- Lysine) tetrachlorocuprate combined with ascorbate and basic curcumin half an hour prior to the administration of Fe-NTA and chloroform till the end of 4 months.

### 2.3. Sampling

At the end of the experiment (4 months), all animals were sacrificed, blood samples were collected and serum was separated by centrifugation at 2500 r.p.m for 15 minutes, the clean clear serum was separated by Pasteur pipette and kept in a deep freeze at -20 °c until used for biochemical analysis. Also, livers tissue specimens of the experimental animal's groups will be quickly removed, perfuse immediately with ice-cold saline (0.9%w/u) and homogenized (Glas-Col, Terr Hauter, USA) in chilled phosphate buffer (0.1 M, pH 7.4) containing KCl (1.17%w/u). The homogenate was centrifuged at 3000 rpm for 10 min at 4°c and clear supernatants were used for biochemical analysis. Additionally, a portion of freshly excised rat liver, of all groups, was dissected and immediately

fixed in 8% phosphate buffered formalin and used for histopathological examination.

### 2.4. Biochemical analysis:

Determination of MPO, MDA, GSH and CAT were analyzed according to the methods described by (Joris *et al.*, 2006), (Ohkawa *et al.*, 1979), (Beutler *et al.*, 1963) and (Aebi, 1984 & Fossati *et al.*, 1980) respectively. Determination of TNF- $\alpha$  and IL-1 using ELISA kit according to the methods described by (Chen *et al.*, 1998) and (Liu *et al.*, 1995) respectively.

### 2.5. Statistical Analysis

All data were expressed as the mean  $\pm$  SD. Statistical comparison between different groups were done by one-way analysis of variance (ANOVA) followed by Tukey's post hoc test for multiple comparison (Graphpad Prism version 5.03, San Diego, CA, USA).  $P < 0.05$  was considered to be statistically significant.

## 3. RESULTS

### 3.1. Effect of basic curcumin, tetrachlorocuprate-lysine and ascorbic acid on the MDA, GSH, MPO and catalase activity:

The obtained data in table (1) revealed that, the mean levels of the MDA of the carcinogen-treated group were (6.22 $\pm$ 0.58), which showed a significant increase compared to the mean levels of the control group (2.35  $\pm$  0.19) at the same period of treatment ( $p \leq 0.05$ ). Whereas the mean levels of the MDA were (3.79 $\pm$ 0.14) in curcumin-treated group, (3.46 $\pm$ 0.12) in tetrachlorocuprate-lysine+ascorbic acid-treated group and (3.12 $\pm$ 0.02) in the mixture-treated group at the same period of treatment ( $p \leq 0.05$ ) which showed a significant decrease compared to the mean levels of the carcinogen-treated group.

The mean levels of the reduced glutathione of the carcinogen-treated group were (14.67 $\pm$ 1.11), which showed a significant decrease compared to the mean levels of the control group (29.37 $\pm$  0.99) at the same period of treatment ( $p \leq 0.05$ ). Whereas the mean levels of the GSH were (24.13 $\pm$ 0.7) in

curcumin-treated group,  $(27.88 \pm 0.61)$  in tetrachlorocuprate-lysine+ascorbic acid-treated group and  $(29.50 \pm 0.60)$  in the mixture-treated group at the same period of treatment ( $p \leq 0.05$ ) which showed a significant increase compared to the mean levels of the carcinogen-treated group, table (1). The obtained data in table (1) showed that, the mean levels of the MPO of the carcinogen-treated group were  $(1.79 \pm 0.07)$ , which indicated a significant elevation compared to the mean levels of the control group  $(0.22 \pm 0.01)$  at the same period of treatment ( $p \leq 0.05$ ). Whereas the mean levels of the MPO were  $(0.84 \pm 0.06)$  in curcumin-treated group,  $(0.82 \pm 0.05)$  in tetrachlorocuprate-lysine+ascorbic acid-treated group and  $(0.82 \pm 0.05)$  in the mixture-treated group at the same period of treatment ( $p \leq 0.05$ ) which resulted in a significant reduction compared to the mean levels of the carcinogen-treated group. The obtained data in table (1) showed that, the mean levels of the catalase of the carcinogen-treated group were  $(70.53 \pm 3.119)$ , which indicated a significant reduction compared to the mean levels of the control group  $(122.5 \pm 1.76)$  at the same period of treatment ( $p \leq 0.05$ ). Whereas the mean levels of the Catalase were  $(96.35 \pm 1.85)$  in curcumin-treated group,  $(101.0 \pm 4.14)$  in tetrachlorocuprate-lysine+ascorbic acid-treated group and  $(111.2 \pm 1.88)$  in the mixture-treated group at the same period of treatment ( $p \leq 0.05$ ) which resulted in a significant reduction compared to the mean levels of the carcinogen-treated group.

### 3.2. Effect of the different compounds on tissue Interleukin-1 and TNF- $\alpha$ activity:

The obtained data in table (2) revealed that, the mean levels of the Interleukin-1 of the carcinogen-treated group were  $(113.5 \pm 1.21)$ , which showed a significant increase compared to the mean levels of the control group  $(30.89 \pm 1.011)$  at the same period of treatment ( $p \leq 0.05$ ). Whereas the mean levels of the Interleukin-1 were  $(86.50 \pm 1.97)$  in curcumin-treated group,  $(77.10 \pm 1.41)$  in tetrachlorocuprate-lysine+ascorbic acid-treated group and

$(61.52 \pm 3.93)$  in the mixture-treated group at the same period of treatment ( $p \leq 0.05$ ) which showed a significant decrease compared to the mean levels of the carcinogen-treated group. The mean levels of the TNF- $\alpha$  of the carcinogen-treated group were  $(117.6 \pm 2.42)$ , which showed a significant increase compared to the mean levels of the control group  $(33.04 \pm 0.97)$  at the same period of treatment ( $p \leq 0.05$ ). Whereas the mean levels of the TNF- $\alpha$  were  $(69.95 \pm 2.29)$  in curcumin-treated group,  $(69.07 \pm 1.18)$  in tetrachlorocuprate-lysine+ascorbic acid-treated group and  $(47.40 \pm 1.71)$  in the mixture-treated group at the same period of treatment ( $p \leq 0.05$ ) which showed a significant decrease compared to the mean levels of the carcinogen-treated group, table (2).

## 4. DISCUSSION

Lipid peroxidation play an important role in carcinogenesis (Banakar et al., 2004), is the most studied biologically free radical chain reaction. Lipid peroxidation may lead to the formation of several toxic byproducts such as malondialdehyde (MDE) and 4-hydroxynonenal which can attack cellular targets including DNA and lead to mutagenicity. An excellent model of in vivo free radical induced damage, associated with extensive lipid peroxidation, is the ferricnitrilotriacetate (Fe-NTA) model. Administration of Fe-NTA leads to increasing oxidative stress that starts from the plasma compartment, where Fe-NTA finds the ideal environment to react with oxidizable lipids (Deiana et al., 2007). This may explain the elevated levels of MDA in rats administrated with Fe-NTA. The obtained results of this work demonstrated that, feeding rats with basic curcumin decreased the lipid peroxidation level compared to the carcinogen group, in agreement with the study of Sankar et al., (2012) who showed that, the presence of curcumin significantly decreased MDA as a marker of lipid peroxidation. Lipid

Table.1) Effect of basic curcumin, tetrachlorocuprate-lysine and ascorbic acid on the serum GSH, MAD and tissue CAT and MPO activities in carcinogen-treated rats:

Parameter Groups	GSH (μmol/ L)	MDA (μmol/ L)	CAT (U/g)	MPO (U/g)
Control	29.37± 0.99 <sup>a</sup>	2.35 ± 0.19 <sup>a</sup>	122.5±1.76 <sup>a</sup>	0.22± 0.01 <sup>a</sup>
Carcinogen	14.67±1.11	6.22±0.58	70.53 ± 3.119	1.79±0.07
Curcumin+ Carcinogen	24.13±0.7 <sup>b</sup>	3.79±0.14 <sup>b</sup>	96.35± 1.8 <sup>b</sup>	0.84±0.06 <sup>bc</sup>
Tetraclorocuprate -lysine+Ascorbic+ Carcinogen	27.88±0.61 <sup>b</sup>	3.46±0.12 <sup>b</sup>	101.0 ± 4.14 <sup>b</sup>	0.82±0.05 <sup>bc</sup>
Mixture+ Carcinogen	29.50±0.60 <sup>b</sup>	3.12±0.02 <sup>b</sup>	111.2± 1.88 <sup>b</sup>	0.37±0.04 <sup>b</sup>

Data are presented as (Mean ± S.E). S.E = Standard error. Mean values with different superscript letters in the same column are significantly different at ( $P<0.05$ ).

Table.2) Effect of basic curcumin, tetrachlorocuprate-lysine and ascorbic acid on the tissue TNF-  $\alpha$  and IL-1 activities in carcinogen-treated rats:

Parameter Groups	IL-1 (pg/ml)	TNF- $\alpha$ (pg/ml)
Control	30.89± 1.011 <sup>a</sup>	33.04±0.97 <sup>a</sup>
Carcinogen	113.5±1.21	117.6 ± 2.42
Curcumin+ Carcinogen	86.50±1.97 <sup>bc</sup>	69.95 ± 2.29 <sup>bc</sup>
Tetraclorocuprate -lysine+Ascorbic+ Carcinogen	77.10±1.41 <sup>bc</sup>	69.07 ± 1.18 <sup>bc</sup>
Mixture+ Carcinogen	61.52±3.93 <sup>b</sup>	47.40 ±1.71 <sup>b</sup>

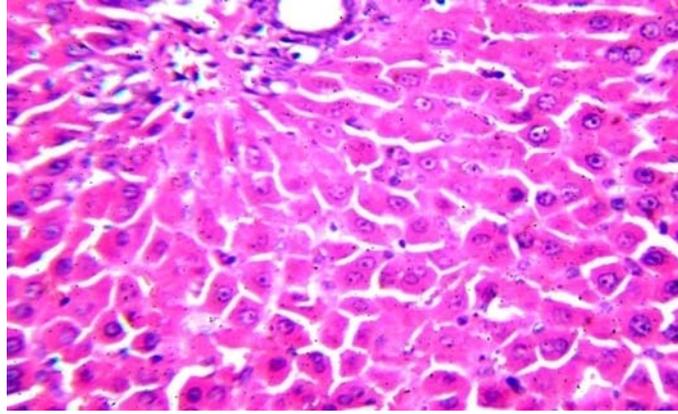


Fig (1) A photomicrograph of section of normal-looking liver of control group showing blond hepatocytes and normal architecture (400 X).

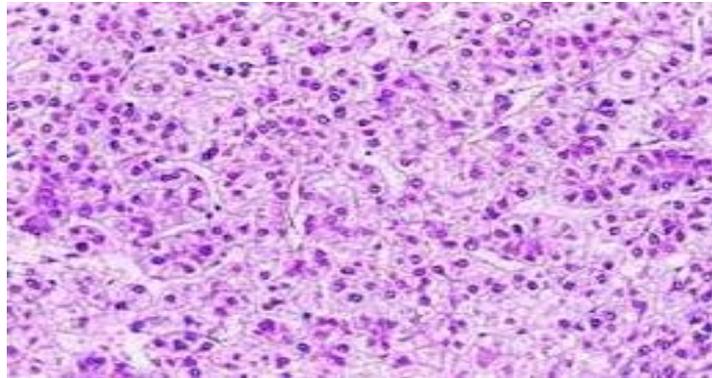


Fig (2) A photomicrograph of section of liver tissue in carcinogenic group showing malignant nuclei (X400).

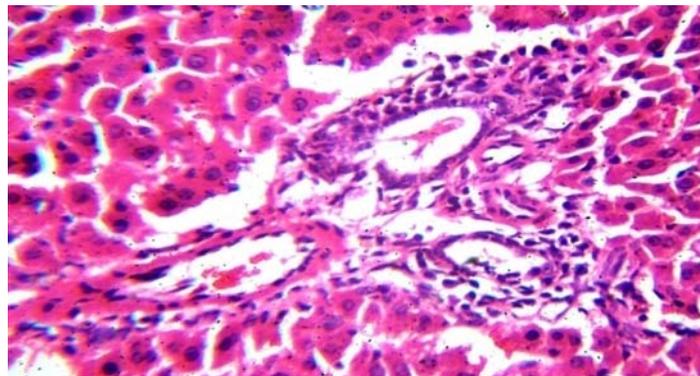


Fig (3) A photomicrograph of section of liver of curcumin treated carcinogenic group showing normal-looking portal tract (X400).

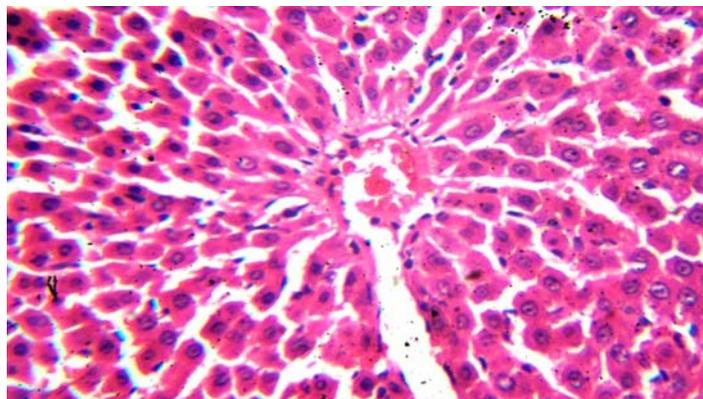


Fig (4) A photomicrograph of section of liver of tetrachlorocuprate-lysine and ascorbic acid treated carcinogenic group showing normal trabecula of hepatocytes and central vein (X400).

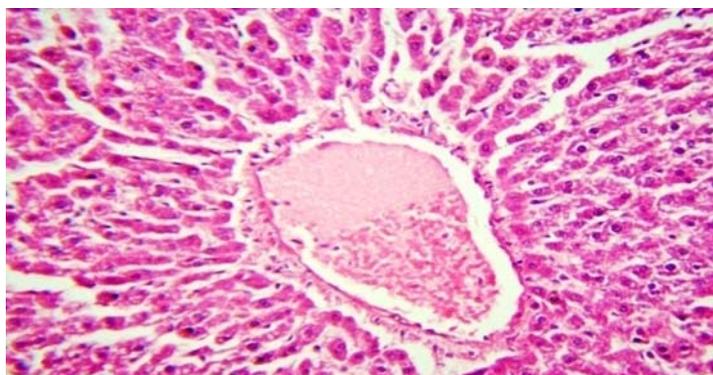


Fig (5) A photomicrograph of section of liver of Curcumin, tetrachlorocuprate-lysine and ascorbic acid treated carcinogenic group showing trabecula of hepatocytes (1-2 cell thick) and central vein (X400).

peroxidation was decreased in the presence of ascorbic acid. This result was in agreement with Elias *et al.*, (2012) who showed that, Vitamin C (ascorbic acid) which is a major water-soluble antioxidant is believed to decrease lipid peroxidation either directly or indirectly by regenerating vitamin E. Vitamin C is an important free radical scavenger in extracellular fluids, trapping radicals and protecting biomembranes from peroxide damage. Malondialdehyde (MDA) concentration was decreased in plasma and liver tissues, by Cu administration (Jianbo *et al.*, 2011). This explains the decreased levels of MDA in rats administrated with tetrachlorocuprate. Glutathione (GSH) plays an important role in a multitude of cellular processes, including cell differentiation, proliferation, and apoptosis, and disturbances in GSH homeostasis are

involved in the etiology and progression of many human diseases including cancer (Nicola *et al.*, 2013). Compared with the control, GSH concentrations in treated rats significantly decreased after exposure to Fe-NTA in agreement with the reports of (Athar & Iqbal., 1998) who showed the decreased levels of GSH in rats administrated with Fe-NTA. The present study showed that administration of basic curcumin increased the GSH levels compared to the carcinogen non treated group. Similarly, Chiagoziem *et al.*, (2014) demonstrated that administration of curcumin improved the GSH. It was clear from the present study that a significant increase in serum GSH levels was induced after administration of tetrachlorocuprate compared with that received Fe-NTA and Chloroform. This improvement in the glutathione levels may

be referred to supplementation with copper increase GSH levels (Derouiche et al., 2013) and presence of ascorbic acid which is important in the production of glutathione (Duke and Atchley, 1984).

Catalase is a very important enzyme in protecting the cell from oxidative damage by reactive oxygen species (Biology in focus, 2008). In the present study, a decrease in liver catalase activity was observed in rats injected with Fe-NTA compared to control group. These results were in agreement with Athar and Iqbal (1998) who showed that the catalase activity was significantly decreased. In our study, the mean values of catalase activity in rats administrated with basic curcumin showed a significant elevation as compared to carcinogen non treated group in agreement with the reports of Prabhakar et al., (2007).

It was clear from the present study that a significant increase in serum catalase activity was induced after administration of ascorbic acid which combined with tetrachlorocuprate and in agreement with the reports of Garg and mahajan., (1993). Myeloperoxidase (MPO) is a peroxidase enzyme, most abundantly expressed in neutrophil granulocytes and produces hypohalous acids to carry out their antimicrobial activity (Klebanoff, 2005).

It was clear from the present study that a significant increase in liver myeloperoxidase activity was induced after administration of Fe-NTA was agreement with the reports of Muneeb et al., (2012).

The present study also showed that, administration of basic curcumin significantly decreased the MPO levels compared to the carcinogen group. Also, Zhe et al., (2014) demonstrated that, treatment with curcumin significantly reduced the expression of MPO.

It was shown that administration of ascorbic acid caused a decrease in MPO activity (Murat et al., 2008).

Interleukins are a group of cytokines (secreted proteins and signal molecules) that were first seen to be expressed by

leukocytes (Brocker et al., 2010). Interleukin-1 alpha and interleukin-1 beta (IL-1 $\alpha$  and IL-1 $\beta$ ) are cytokines that participate in the regulation of immune responses, inflammatory reactions, and hematopoiesis (Barthelmes et al., 2011). It is obvious from the present study that Interleukin-1 activity in rats administrated with Fe-NTA and Chloroform was significantly elevated compared with its corresponding values in normal group and in agreement with the reports of Ahmad et al., (2005). The results of this work demonstrated that, feeding rats with basic curcumin decreased the Interleukin-1 activity compared to the carcinogen group. Moreover, Gaddipati et al., (2003) reported that, oral administration of curcumin resulted in significant restoration of the cytokines included interleukin-1 to depleted levels.

It was clear from the present study that, a significant decrease in liver interleukin-1 levels was observed after administration of ascorbic acid when compared to carcinogen group. The obtained results were nearly similar to Tomofuji et al., (2009) who demonstrated that, gene expression encoding inflammation, including interleukin-1 alpha and interleukin-1 beta, was more than two fold down-regulated by vitamin C intake.

Tumor necrosis factor (TNF) is a multifunctional cytokine that plays important roles in diverse cellular events such as cell survival, proliferation, differentiation, and death. As a pro-inflammatory cytokine, TNF is secreted by inflammatory cells, which may be involved in inflammation-associated carcinogenesis. Recent studies have focused on sensitizing cancer cells to TNF-induced apoptosis through inhibiting survival signals such as NF- $\kappa$ B, by combined therapy (Xia & Yong., 2008). In addition to apoptosis, TNF can also induce necrotic cell death. Reactive oxygen species (ROS) play a critical role in mediating necrotic cell death because ROS scavenger butylated hydroxyanisole (BHA)

can effectively block this pathway (Lin *et al.*, 2004).

The present study showed that administration of Fe-NTA increased the TNF- $\alpha$  levels compared to the control group. Similarly, Summya *et al.*, (2013) demonstrated that chronic exposure of Fe-NTA for 16 weeks in tumor group induced over expression of TNF- $\alpha$  as compared to control group. Also, Muneeb *et al.*, (2012) reported that, elevated levels of TNF- $\alpha$  could be detected in serum of rats exposed to tumor promotion with Fe-NTA. Compared with the carcinogen group, TNF- $\alpha$  concentration in treated rats significantly decreased after administration of curcumin which had been shown to markedly reduce serum TNF- $\alpha$  (Gulcubuk *et al.*, 2006). This explains the decreased levels of TNF- $\alpha$  in rats administrated with basic curcumin.

In the present work, a significant decrease in liver TNF- $\alpha$  level was observed in rats administrated with ascorbic acid compared with carcinogen group. Also, Masoumeh *et al.*, (2015) revealed that, ascorbic acid administration could reduce the levels of TNF- $\alpha$ . Moreover, Senturk *et al.*, (2004) demonstrated that, ascorbic acid administration significantly decreased the concentrations of serum tumor necrosis factor- $\alpha$ . It was clear from the present study that a significant decrease in liver TNF- $\alpha$  levels was induced by administration of copper-lysine. Also, Jianbo *et al.*, (2008) demonstrated that, the reduction in backfat depth may be due to copper from Cu-lysine altering TNF- $\alpha$  metabolism in lambs.

Histopathological examination of the liver tissues of the different experimental groups illustrated that, Fe-NTA and chloroform administrated group showed macrovesicular steatosis with prominent sinusoid nucleoli. Hepatic arteriole showed Hyalinosis. dilated central venule as well as wide area of coagulative necrosis and malignant nuclei compared to that of control group. These results agree with Kannappan (2014) who reported that, an iron chelate, ferric nitrilotriacetate (Fe-NTA), induces necrosis as a consequence of

lipid peroxidation and oxidative tissue damage that eventually leads to high incidence of cancer in liver and kidney. Fe-NTA acts through the generation of free radicals and by enhancing the rate of DNA synthesis with simultaneous decrease in antioxidant defenses. Also, Chloroform may be caused enhancement of cancer induction in was in agreement with Byron *et al.*, (1995) who showed that, Chloroform produces cancer by a non genotoxic-cytotoxic mode of action, with no increased cancer risk expected at noncytotoxic doses. Rat's liver administrated with curcumin revealed that, significant improvement in the histological structure of liver tissue compared to the carcinogen group. Similarly, Ajaikumar *et al.*, (2008) reported that, curcumin has been found to inhibit the proliferation of various tumor cells in culture, prevents carcinogen-induced cancers in rodents, and inhibits the growth of human tumors in xenotransplant or orthotransplant animal models either alone or in combination with chemotherapeutic agents. Also, the results have indicated that, tetrachlorocuprate-lysine in combination with ascorbic acid administration showed significant improvement in the histological picture compared to the carcinogen group. Also, Netke *et al.*, (2003) reported that the synergistic anticancer effect of ascorbic acid, proline, lysine on several cancer cell lines in tissue culture studies was greater than that of the individual nutrients. Moreover, Casini *et al.*, (2007) reported that, the interactions with protein targets of the ruthenium(III) complex imidazolium trans- [tetrachloro(dimethyl sulfoxide) (imidazole) ruthenate(III)], NAMI-A, an effective anticancer and anti-metastatic agent now in clinical trials.

**Conclusion:** Basic curcumin has potent chemopreventative activity against a wide variety of tumors and has great potential in the prevention and treatment of hepatocarcinogenesis, Moreover, Tetrachlorocuprate- lysine in combination with ascorbic acid exert chemopreventative effect against hepatocellular-carcinoma,

through have antioxidant and free radicals scavenging activity and trapping of activated metabolites of chemical carcinogen.

## 5. REFERENCES

- Adams, C.J., Haddow M.F., Hughes R. J.; Kurawa, M.A. and Orpen G.A. 2010. Dalton Trans., 39: 3714.
- Adejuwon, A.A. and Joseph, O.O. 2008. Protective Effect of Oral Ascorbic Acid (Vitamin C) Against Acetaminophen- Induced Hepatic Injury in Rats. African Journal of Biomedical Research, 11: 183-190.
- Aebi, H. 1984. Catalase in vitro. Methods Enzymol, 105:121–126.
- Afrah, F.S ; Ehab, T.; Kamal, A.F.S. and Hind, T. H. 2014. Protective effect of curcumin on chloroform as by-product of water chlorination induced cardiotoxicity. Biomedicine and Preventive Nutrition, 4(2): 225-230.
- Agency for Toxic Substances and Disease Registry (ATSDR), 1997. Toxicological Profile for Chloroform. Public Health Service, U.S. Department of Health and Human Services, Atlanta, GA., 36:170.
- Ahmad, N.S., Khalid, B.A., Luke, D.A. and Ima, Nirwana, S. 2005. Tocotrienol offers better protection than tocopherol from free radical-induced damage of rat bone. Clin Exp Pharmacol Physiol., 32(9):761-70.
- Ajaikumar, B.K.; Preetha, A. and Bharat, B.A. 2008. Curcumin inhibits proliferation, invasion, angiogenesis and metastasis of different cancers through interaction with multiple cell signaling proteins, 8:199–225.
- Akram, M.; Shahab, U.; Afzal, A.; Khan, U.; Abdul, H.; Mohiuddin, E. and Asif, M. 2010. CURCUMA LONGA AND CURCUMIN: A REVIEW ARTICLE. ROM. J. BIOL. – PLANT BIOL., 55(2): 65–70.
- Athar, M. and Iqbal, M. 1998. Ferric nitrotriacetate promotes N-diethylnitrosamine-induced renal tumorigenesis in the rat: implications for the involvement of oxidative stress. Carcinogenesis, 19(6):1133-9.
- Awai, M., Narasaki, M., Yamanoi, Y. and Seno, S. 1979. "Induction of diabetes in animals by parenteral administration of ferric nitrotriacetate. A model of experimental hemochromatosis," The American Journal of Pathology, 95: 663–673.
- Banakar, M.C.; Paramasivan, S.K. ; Chattopadhyay, M.B.; Datta, S. P. ; Barthelmes, K.; Reynolds, A.M.; Peisach, E.; Jonker, H.R.; DeNunzio, N.J.; Allen, K.N.; Imperiali, B. and Schwalbe, H. 2011: "Engineering encodable lanthanide-binding tags into loop regions of proteins". J. Am. Chem. Soc. 133 (4): 808–19.
- Barthelmes, K.; Reynolds, A.M. ; Peisach, E.; Jonker, H.R.; DeNunzio, N.J.; Allen, K.N. ; Imperiali, B. and Schwalbe, H. 2011. "Engineering encodable lanthanide-binding tags into loop regions of proteins". J. Am. Chem. Soc. 133 (4): 808–19.
- Beutler, E. O. and Kelly, B.M. 1963. Improved method for the determination of blood glutathione. J. Lab. Clin. Med., 61: 882-888.
- Biology in focus (2008): HSC course / Glenda Chidrawi, Margaret Robson with Stephanie Hollis, 8:119
- Brocker, C.; Thompson, D.; Matsumoto, A.; Nebert, D.W. and Vasiliou, V. 2010. "Evolutionary divergence and functions of the human interleukin (IL) gene family." Human Genomics, 5 (1): 30–55.
- Byron, E. B.; Michael, V. T.; Rory, B. C.; Gregory, L. K. and Douglas, C. W. 1995. The role of regenerative cell proliferation in chloroform-induced cancer. 82: 23–26.
- Casini, A.; Mastrobuoni, G.; Terenghi, M; Gabbiani, C.; Monzani, E.; Moneti, G.; Casella, L. L. and Messori, L. 2007. A study of the interactions of the ruthenium(III) complex

- imidazolium trans-[tetrachloro(dimethyl sulfoxide)(imidazole)ruthenate(III)] with hen egg white lysozyme and horse heart cytochrome c. *Biol Inorg Chem*, 12(8):1107-17.
- Chen, W.; Jin, W.; Cook, M.; Weiner, L. H. and Wahl, M. S. 1998. Oral delivery of group A streptococcal cell walls augments circulating TGF-beta and suppresses streptococcal cell wall arthritis. *J. Immunol.*, 161: 6297-6304.
- Chiagoziem, A. O; Sunny, O. A; Victoria, I. O; Azeezat, L. A, and Oluwafemi, E. K. 2014. Protective Effect of Curcumin against the Liver Toxicity Cause Prophylactic Effect of Vitamin C on Cyclosporine A-induced Liver Toxicity by Propanil in Rats, 210(8): 177-185.
- Daniel, K.G. ; Chen, D. ; Orlu, S. ; Cui, Q.C. ; Miller, F.R. and Dou, Q.P. 2005. "Clioquinol and pyrrolidine dithiocarbamate complex with copper to form proteasome inhibitors and apoptosis inducers in human breast cancer cells". *Breast Cancer Res.* 7 (6): 897-908.
- Deiana, M.; Rosa, A.; Corona, G.; Atzeri, A.; Incani, A.; Visioli, F. ;Melis, M.P. and Dessi, M.A. 2007. Protective effect of olive oil minor polar components against oxidative damage in rats treated with ferric-nitritolriacetate. *Food Chem Toxicol*, 45: 2434-2440.
- Derouiche, S.; Kawther, A.; Manel, D.; Soumya, B.A. and Kechrid, Z. 2013. The effects of copper supplement on zinc status, enzymes of zinc activities and antioxidant status in alloxan-induced diabetic rats fed on zinc overdose diet. *International Journal of Nutrition and Metabolism*, 5(5): 82-87.
- Duke, J.A. and Atchley, A.A. 1984: Proximate analysis. In: Christie, B.R. (Ed.), the Handbook of Plant Science in Agriculture. CRC Press, Inc., Boca Raton, Florida, 32: 60
- Elias, Adikwu and Oputiri Deo. 2013: Hepatoprotective Effect of Vitamin C (Ascorbic Acid). *Pharmacology & Pharmacy*, 4, 84-92.
- Fossati, P.; Prencipe, L. and Berti, G. 1980. Use of 3, 5-dichloro-2-hydroxybenzenesulfonic acid/4-aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. *Clin Chem.*, 26(2): 227-31.
- Frechilla, D.; Lasheras, B.; Ucelay, M; Parrondo, E.; Craciunescu, G. and Cenarruzabeitia, E. 1990. On the mechanism of the anti-inflammatory activity of some copper (II) complexes. *Arzneimittelforschung*, 40(9):1008-10.
- Gaddipati, J.P.; Sundar, S.V.; Calemine, J.; Seth, P.; Sidhu, G.S ; Maheshwari, R.K. 2003: Differential regulation of cytokines and transcription factors in liver by curcumin following hemorrhage/resuscitation. *Shock.*, 19(2):150-6.
- Garg, S. K. and Mahajan, S. 1993: Effect of ascorbic acid on longevity, catalase and lipid peroxidation in *Callosobruchus maculatus* F. Springer International Publishing AG, 16(3):87-92.
- Gulcubuk, A.; Altunatmaz, K.; Sonmez, K.; Haktanir-Yatkin, D. ; Uzun, H. ; Gurel, A. and Aydin, S. 2006. Effects of curcumin on tumour necrosis factor-alpha and interleukin-6 in the late phase of experimental acute pancreatitis. *J Vet Med A Physiol Pathol Clin Med.*, 53(1):49-54.
- Iqbal, M.; Giri, U.; Giri, D.K.; Alam, M.S. and Athar, M. 1999. Age-dependent renal accumulation of 4-hydroxy-2-nonenal (HNE)-modified proteins following parenteral administration of ferric nitritolriacetate commensurate with its differential toxicity: Implications for the involvement of HNE-protein adducts in oxidative stress and

- carcinogenesis. *Arch Biochem Biophys.*, 365:101–12.
- Jianbo, C; Hui, M.; Caiyun, F.; Zijun, Z.; Zhihai, J.; Xiaoping, Z. and Lisheng, W. 2011. Effects of Different Copper Sources and Levels on Plasma Superoxide Dismutase, Lipid Peroxidation, and Copper Status of Lambs. *Biological Trace Element Research*, 144 (1): 570-579.
- Joris, J.T.H. R.; Kasper, M.A. R.; Jaklien, C. L.; Nike, C.; Anita M. de B.; Wilma, M. F.; Lijnen H. R.; Jan, J. W. and Sandrine, F. 2006. Tissue-Type Plasminogen Activator Modulates Inflammatory Responses and Renal Function in Ischemia Reperfusion Injury. *Journal of the American society of nephrology*, 17(1): 131-140.
- Kannappan, P.; Palanisamy, C. P. and Velliyur, K., G.. 2014. Protective Effect of Ethanolic Extract of *Tabernaemontana divaricata* (L.) R. Br. against DEN and Fe NTA Induced Liver Necrosis in Wistar Albino Rats. *Biomed Res Int.*, 20: 240-243.
- Kaur, G.; Lone, I.; Athar, M. and Alam, M.S. 2007. Protective effect of *Didymocarpus pedicellata* on ferric nitrilotriacetate (Fe-NTA) induced renal oxidative stress and hyperproliferative response. *Chem Biol Interact.*, 165:33–44.
- Klasing, K. C. 1998. Minerals. In *Comparative Avian Nutrition*. CAB international. New York. USA, 18. 234-276.
- Klebanoff, S. J. 2005. Myeloperoxidase: friend and foe. *J. Leukoc. Biol.* 77, 598–625.
- Kumar, V, Fausto N, Abbas, A. 2003. *Robbins & Cotran Pathologic Basis of Disease* (7th ed.). Saunders., 31:914–927.
- Lachapelle, M. Y. and Drouin, G. 2010. "Inactivation dates of the human and guinea pig vitamin C genes". *Genetica*, 139 (2): 199–207.
- Lin, Y.; Choksi S.; Shen, H.M.; Yang, Q.F.; Hur, G.M. and Kim, Y.S. 2004. Tumor necrosis factor-induced nonapoptotic cell death requires receptor-interacting protein-mediated cellular reactive oxygen species accumulation. *J Biol Chem.*, 279(11):10822–8.
- Liu, X.; Yang, J. and Wang, S., 1995. Interleukin-1 beta, interleukin-1 receptor antagonist mRNA expression of peripheral blood mononuclear cells (PMNC) in idiopathic nephritic syndrome detected by biotin-labelled probe in situ hybridization. *Zhonghua Yi Xue Za Zhi.*, 75(3): 152-154.
- Masoumeh, K.; Seyed, M.; Bagher, T.; Katayoun, N.; Jafar, M., S.; Soheila, M.; and Mohammad, B. K. 2015. Evaluation of the Effect of Ascorbic Acid Administration on Gene Expression Level of IL-6 and TNF- $\alpha$  Cytokines in Deceased Donors. *Iran J Allergy Asthma Immunol*, 14(2):149-157.
- Muneeb, U. R.; Mir T. F.; Wajhul, Q.; Rehan, K.; Abdul, Q. K.; Abdul L. ; Oday, O. H. and Sarwat S. 2012. Chemopreventive effect of *Quercus infectoria* against chemically induced renal toxicity and carcinogenesis. *International Journal of Drug Development and Research*, 4(2): 336-351.
- Murat, K.; Halit, I. and Fehmi, O. 2008. Gastroprotective and Anti-oxidative Properties of Ascorbic Acid on Indomethacin-induced Gastric Injuries in Rats. *Biological trace element research*, 126(1-3):222-36.
- Netke, S.P.; Roomi, M.W.; Ivanov, V.; Niedzwiecki, A. and Rath, M. 2003. A Specific Combination Of Ascorbic Acid, Lysine, Proline And Epigallocatechin Gallate Inhibits Proliferation And Extracellular Matrix Invasion Of Various Human Cancer Cell Lines. *Research Communications in Pharmacology*

- and Toxicology: Emerging Drugs, 2: 37-50.
- Nicola, T.; Roberta, R.; Mariapaola, N.; Barbara, M.; Anna, L.; Furfaro, M.; Adelaide, P.; Umberto, M. and Cinzia, D. 2013. Role of Glutathione in Cancer Progression and Chemoresistance. *Oxidative Medicine and Cellular Longevity*, 213:10.
- Ohkawa, H. N. Ohishi and K. Y. agi 1979. Assay for lipid peroxidation in animal tissue by thiobarbituric acid reaction. *Anal. Biochem.*, 95(2): 351-358.
- Prabhakar, R. K.; Veeresh, P. V.; Amit, K.; Beena, M.; Atanu B.; Indira, K. P. and Unnikrishnan K. M. 2007. Effect of Curcumin and Curcumin Copper Complex (1:1) on Radiation-induced Changes of Anti-oxidant Enzymes Levels in the Livers of Swiss Albino Mice, *J. Radiat. Res.*, 48: 241-245.
- Rossberg, M., Lendle, W., Pfeleiderer, G., Tögel, A., Dreher, E.-L., Langer, E. 2006. Chlorinated hydrocarbons. In *Ullmann's Encyclopedia of Industrial Chemistry* (eds). Weinheim, Germany: Wiley-VCH, 26:12.
- Sankar, P.; Telang, A.G. and Manimaran, A. 2012. Protective effect of curcumin on cypermethrin-induced oxidative stress in Wistar rats. *Exp Toxicol Pathol.*, 64(5):487-93.
- Senturk, N.; Keles, G.C.; Kaymaz, F.F.; Yildiz, L.; Acikgoz, G. and Turanli, A.Y. 2004. The role of ascorbic acid on collagen structure and levels of serum interleukin-6 and tumour necrosis factor-alpha in experimental lathyrism. *Clin Exp Dermatol*, 29(2):168-75.
- Summya R.; Nemat, A.; Sana, N.; Syed, K. H. and Sarwat, S. 2013. Amelioration of Renal Carcinogenesis by Bee Propolis: A Chemo Preventive Approach. *Toxicol Int.*, 20(3): 227-234.
- Tomofuji, T.; Ekuni D.; Sanbe, T.; Irie, K.; Azuma, T.; Maruyama, T.; Tamaki, N.; Murakami, J.; Kokeguchi, S. and Yamamoto, T. 2009. Effects of vitamin C intake on gingival oxidative stress in rat periodontitis. *Free Radic Biol Med.*, 46(2):163-168.
- Xia, W. and Yong, L. 2008: Tumor necrosis factor and cancer, buddies or foes?. *Acta Pharmacol Sin.*; 29(11): 1275-1288.
- Yumei, F.; Shizhong, Z.; Jianguo, L.; Jan, R. and Anping, C. 2008. Curcumin protects the rat liver from CCl<sub>4</sub>-caused injury and fibrogenesis by attenuating oxidative stress and suppressing inflammation. *Mol Pharmacol*, 73(2):399-409.
- Zhe, F.; Huirong, J.; Jihong, Y.; Yang, L.; Xiaowei, H.; Huizhu, S.; Gang, S.; Jiyong, P.; Fuwen, L.; and Xiaofeng, T. 2014. The Protective Effects of Curcumin on Experimental Acute Liver Lesion Induced by Intestinal Ischemia-Reperfusion through Inhibiting the Pathway of NF- $\kappa$ B in a Rat Model, 204:8.